

Developing an Oxygen Detection Device for a Microfluidic-based Hypoxia Chamber



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Problem Statement

- Need to understand impact of hypoxic stress on cells
- Use microfluidic devices to generate hypoxic environments
- Will be used to study:
 - Oxidative stress
 - Ischemia
 - Reactive oxygen species – mediated cellular pathways
- Previous semester's work:
 - Produced a functioning microfluidic-based hypoxia chamber
- **This semester's focus:**
 - Develop accurate oxygen detection mechanism for the device

Background

- Heart attacks [1]
 - Kill 600,000 people each year
 - Responsible for 1 in 4 deaths
 - Cardiac cell apoptosis = cell death
- Stem cell fusion to produce new cells
 - Proposed treatment for heart attack patients
- Cell Fusion more likely under hypoxic conditions [2]
- Hypoxic conditions mimicked in microfluidic devices

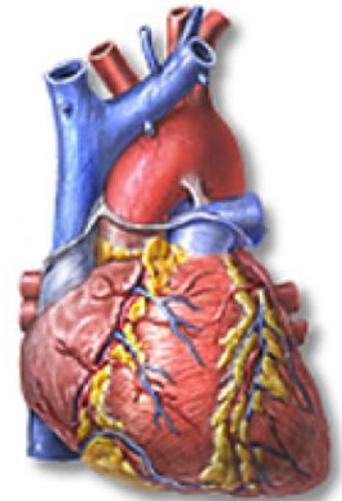


Figure 1. Image of the human heart [1].

Background

- Microfluidics
 - Micro-scale fluid mechanisms
 - Small devices with channels
 - Commonly used with cells
- Ogle Lab Device
 - One time use
 - Made of highly gas permeable poly(dimethylsiloxane) (PDMS)
- Oxygen Sensing/Detection
 - Fluorescent or Luminescent indicators
 - Light source to excite the dye
 - Brightness determines oxygen content



Figure 2. Master slide of microfluidic device developed.

Current Devices

- Commercial devices
 - None for oxygen detection in microfluidic devices
 - General oxygen detection devices
- Research institute devices
 - Oxygen detection methods for specific microfluidic devices
 - Designed specifically for those labs

Commercial Devices

- General oxygen detection

- Thin-film sensors
 - Limited variety in luminescent material
 - Very high cost

- Electrodes
 - Consume oxygen during detection
 - Poor accuracy

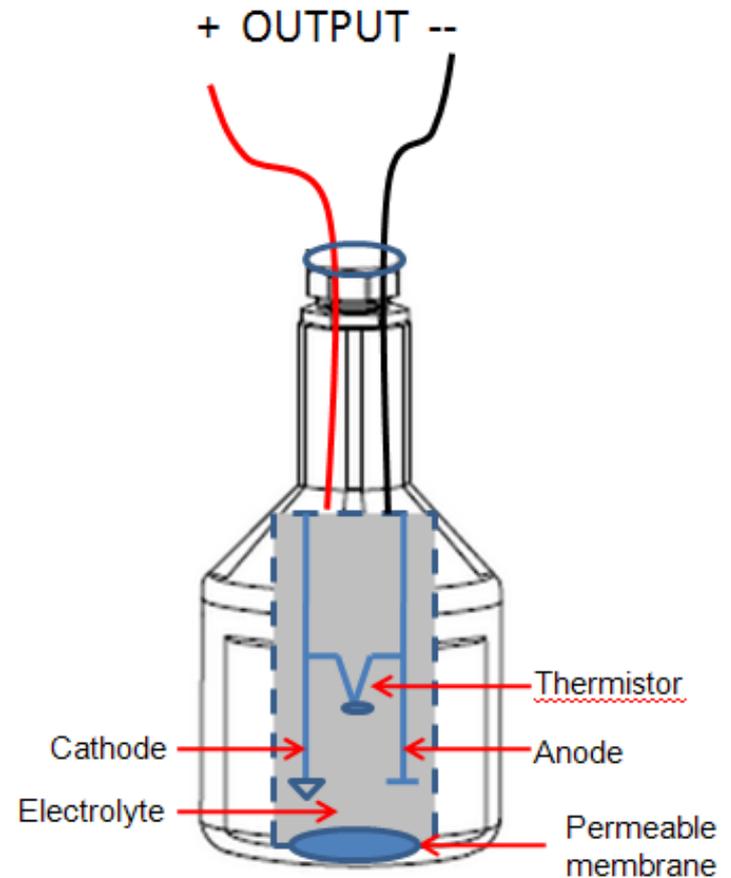


Figure 3. DO6400 Series Dissolved Oxygen Sensor with NI Wireless Sensor Networks (WSN) provided by National Instruments [3].

Research Devices

- Methods include:
 - Thin-film sensors
 - Micro/nanoparticles
 - Water soluble/macroparticles
- Various indicators utilized
- Detection methods:
 - Intensity
 - Fluorescence intensity proportional to concentration
 - Lifetime
 - Exponential decay rate of the fluorescence

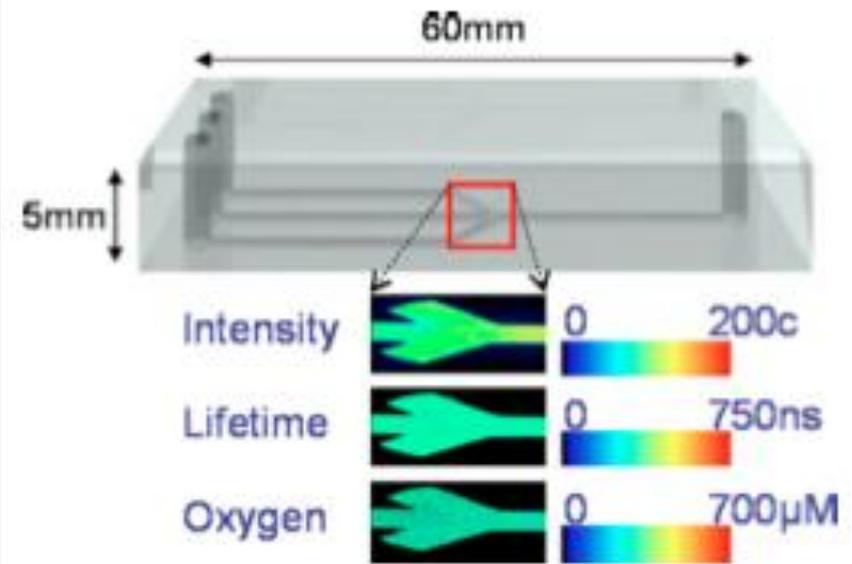


Figure 4. Illustration of fluorescence intensity and lifetime imaging in microfluidic devices using the method developed at University of Michigan [4].

Product Design Specifications

- *Performance Requirements*

- Detect oxygen concentrations from 1% - 21% O₂
- Ability to be used frequently with high level of repeatability

- *Accuracy and Reliability*

Function within a range of +/- 2 to 3% oxygen concentration

- *Life in Service/Shelf Life*

- Last through one experiment (no longer than two weeks)

- *Operation Environment*

- Incubator environment (37°C and 5% CO₂)
- Fluorescent exposure

- *Ergonomics*

- *Low cost*

Design Alternative 1: Thin-film Sensors

- Solution of indicator and encapsulation medium
- Fabricated by pipetting or spinning solution
- Placed directly above or below devices
- Successful with cell culture media
- Widely used already

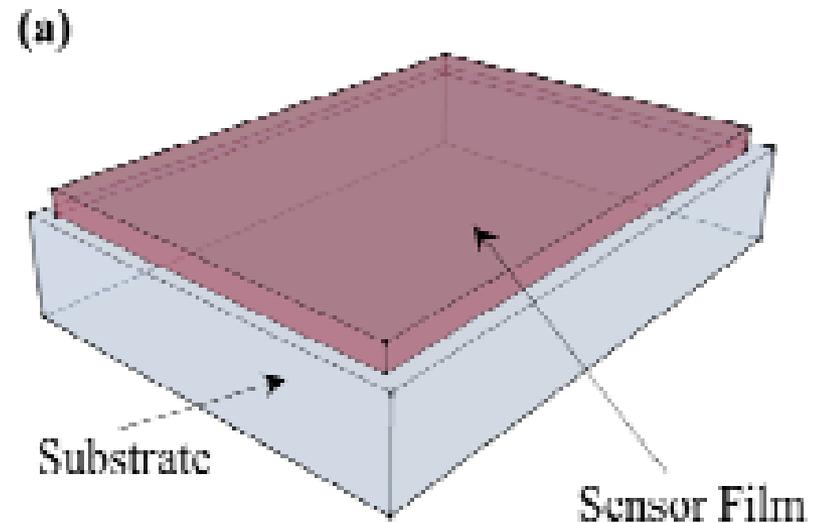


Figure 5. A single thin film sensor on a generic substrate [5].

2: Microparticles/Nanoparticles

- Encapsulated into polymer sensor
- Silica beads doped in indicator
- Added directly to thin films within channels
- High accuracy
- High cost
- Time-consuming

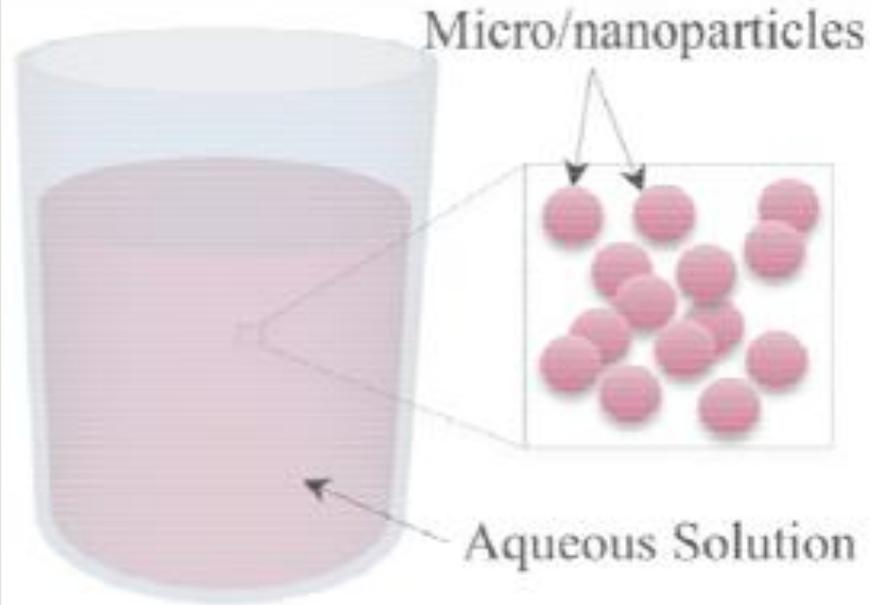


Figure 6. Diagram of micro/nanoparticle sensors suspended in aqueous media [5].

3: Water-soluble Macroparticles

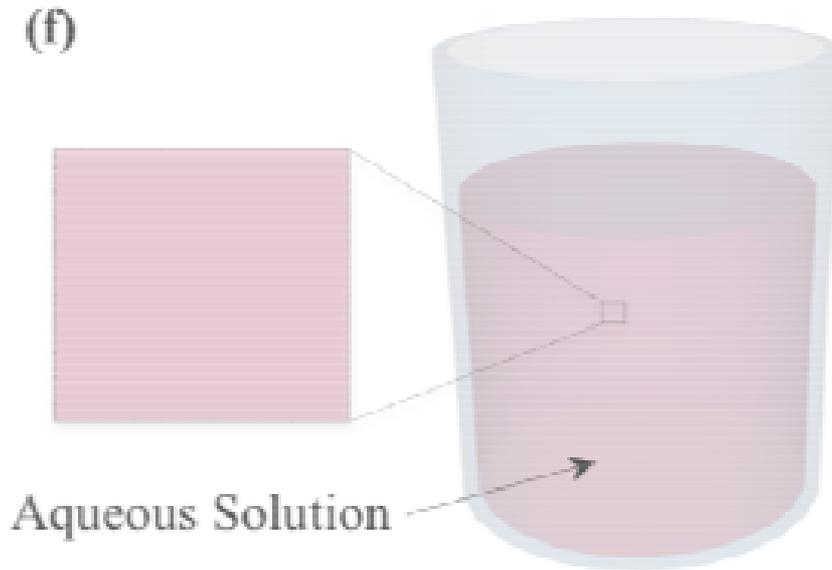


Figure 7. Diagram of water-soluble sensor compound dissolved in aqueous media [5]

- Higher cost
- Improved sensitivity
- Likely to interfere with environment
- Large potential leaching effects
- Time-consuming
- Versatile uses

Design Matrix – Sensor Format

Factors	Thin-Film	Micro/ Nanoparticles	Water-soluble Macroparticles
Accuracy (30)	4	5	2
Cost (25)	3	3	1
Ease of Use (20)	5	4	3
Ease of Assembly (15)	4	3	4
Biocompatibility (10)	5	4	2
Total Points (100)	81	78	45

Indicator Alternative 1: Ruthenium-based

- Very photostable
- Possible cytotoxic effects
 - After repeated excitation
- Lower sensitivity to oxygen
 - Not good for hypoxic conditions
- Used in thin films and nanoparticles

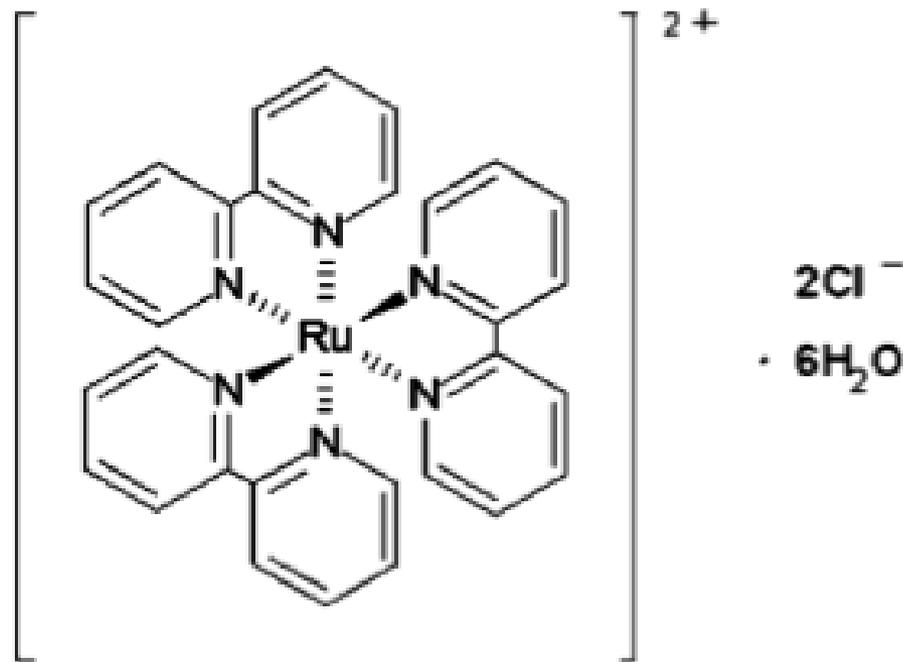


Figure 8. Tris(2,20-bipyridyl dichlororuthenium) hexahydrate, a common ruthenium compound used in optical oxygen sensors [6].

Indicator Alternative 2: Metalloporphyrin-based

- High sensitivity to oxygen
 - Applicable in low-oxygen environments
- Poor photostability
 - PtOEPK and PdOEPK have improved photostability
- No leaching effects

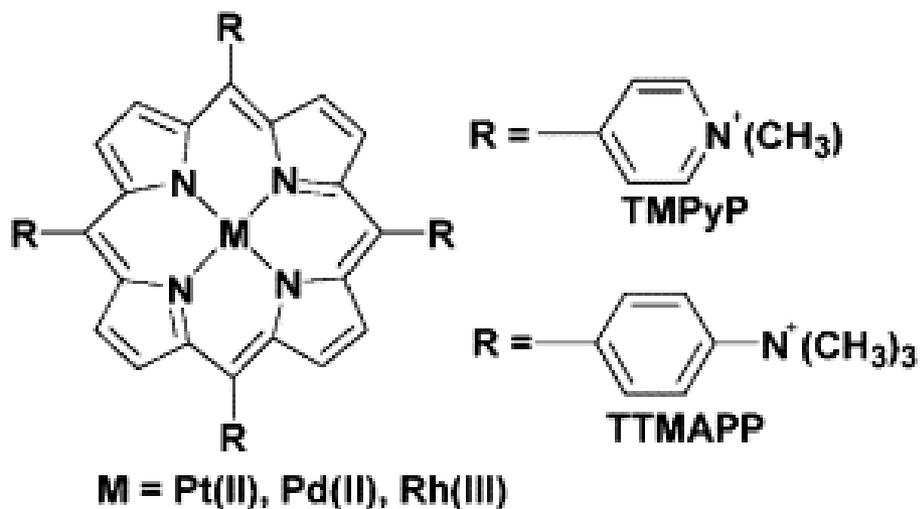


Figure 9. Structures of water-soluble cationic metalloporphyrins [7].

Design Matrix - Indicators

Factors	Ruthenium-based	Metalloporphyrin-based
Detection properties (25)	5	3
Sensitivity to oxygen (30)	2	5
Unquenched Lifetime (10)	2	4
Cost (25)	4	2
Biocompatibility (10)	3	5
Total Points (100)	67	73

Design Selection

- Metalloporphyrin-based indicator
 - PdOEPK or PtOEPK
- Used successfully in optical oxygen sensors
- Increases photostability [5]
- Phosphoresce rather than fluoresce
- Pd exhibits pro-oxidative actions and photo-oxidation [8]
 - Reduced electron density of porphyrin ring

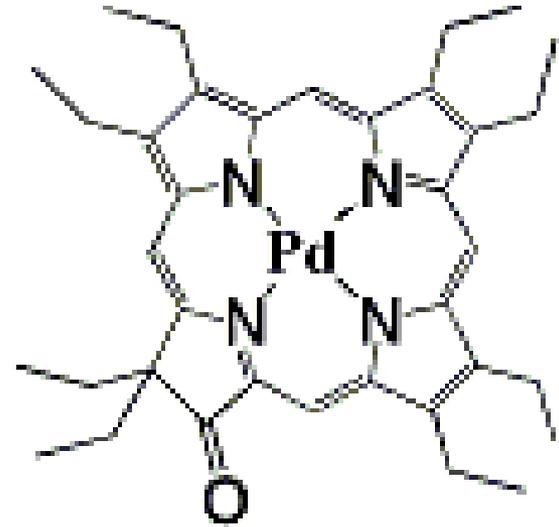


Figure 10. PdOEPK molecule [9].

Design Selection

- Thin film sensor
 - Manufacture with purchased chemicals
 - Made directly onto glass slides
- Encapsulation medium
 - Polystyrene

IMAGE KEY:

-  - PDMS
-  - PdOEPK in Polystyrene
-  - Glass Slide

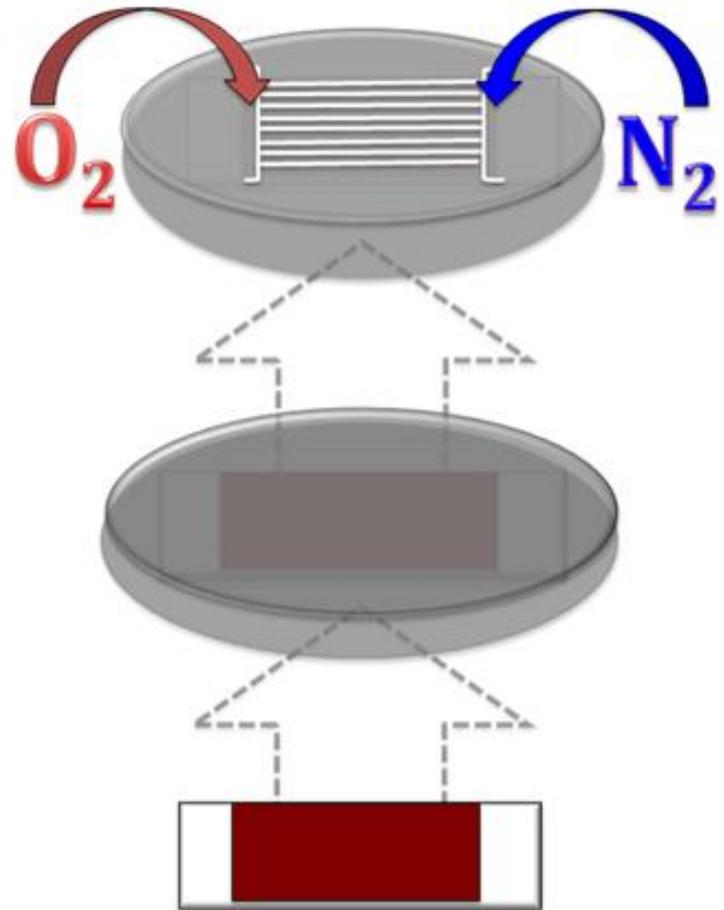


Figure 11. Thin-film oxygen sensor fabricated on a glass slide and placed beneath the microfluidic device for oxygen detection

Future Work

- Simplify oxygen detection system
- Disregard cell media
- Test oxygen sensor apart from the microfluidic device
- Create standardized curve of oxygen concentration

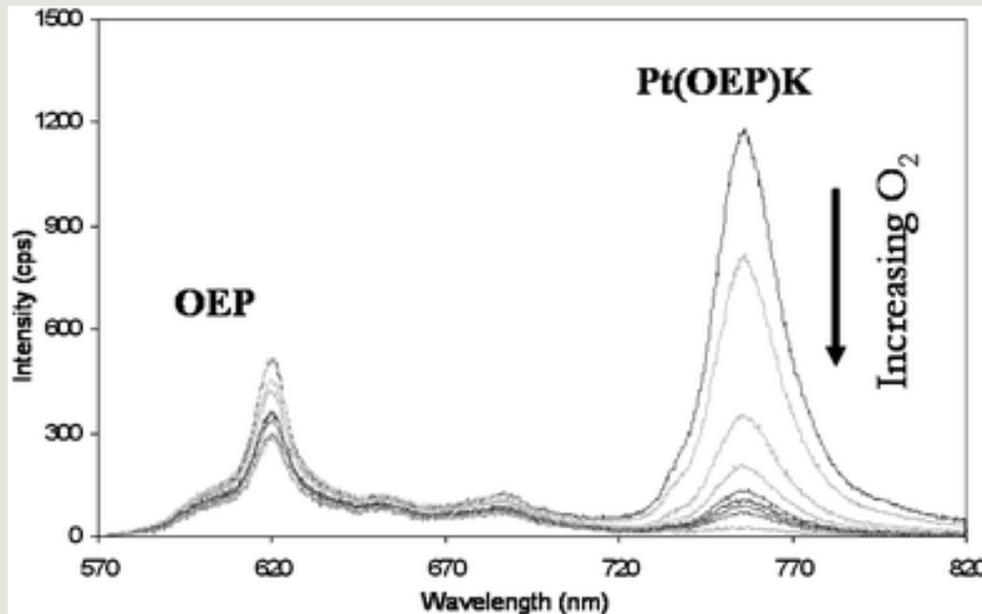


Figure 12. Example of a standardized curve for fiber optic oxygen sensing in various dissolved oxygen concentrations [10].

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Questions



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