



Improving Diagnostic Technology of Acute Compartment Syndrome by Quantifying Intramuscular Glucose Team: Heather Barnwell, Will Bacon, Kristina Geiger, Alexander Goodman, Carly Rogers **Client:** Dr. Christopher Doro **Advisor:** Professor Walter Block

Abstract

Acute compartment syndrome (ACS) is a condition in which the pressure of the muscle compartment increases significantly due to a traumatic injury. The consequence is a decrease in the pressure gradient of blood flow which causes arterial blood to bypass the injured muscle. This trauma leads to cell anoxia, muscle ischemia and muscle death. Diagnosing ACS is problematic as current methods rely on subjective assessments, such as clinical examinations, and uninformative intracompartmental pressure readings. These assessments often return false-positive diagnoses. It has been shown that alternative biochemical markers, specifically glucose, lead to a more indicative quantification of this trauma. While current technology exists to detect glucose in canine models, these are unfit for humans. Our goal is to translate these tools to a clinical setting with a diagnostic glucose probe to analyze the glucose concentration in muscle and increase the specificity of the diagnosis.



Intracompartmental pressure readings are too variable between patients for precise ACS diagnosis. A definitive measure is critical so patients avoid unnecessary and invasive surgeries.^{1,2}

Intramuscular Glucose Detection - Glucose shown to decrease in canines with ACS³ - Sub threshold is a clear indicator of muscle ischemia Translating results to emergency room requires: . Rapid device calibration Time Since Compartment Syndrome Creation (min) 2. Bioelectrochemical sensor Figure 1: Body glucose levels clear digress after onset of acute compartment syndrome. Significant (p = 0.2) glucose 3. Deep tissue penetration (2-8 cm) deviation was detected after 15 minutes and remained stable at 60 (mg/dL).Suggests ACS detection must be done immediately upon E.R. arrival³, Gluconolactone Glucose Biceps femoris (short head) *Figure 2:* The tip of an ACS glucose probe is chemically active

and reacting with glucose in vivo. A redox reaction between glucose, glucose oxidase (GO_v) metal mediator(M_{red}/M_{Ox}) and electrode are shown above. Electron (e) transfer drives current through circuit. More importantly, the reaction is oxygen independent and occurs regardless of compartmental conditions⁴.

Figure 3: ACS probe will enter physical environments such as the anterior muscle compartments of the thigh, shown above. Sensor must reach compartments 3 - 8 centimeters below skin and remain *in vivo* for multiple hours. Scale bars assume a male thigh with a diameter of 15 centimeters⁵.

Proposed Biomarker Mode Diagnostic Accuracy Clinical Treatment







Electrode Engineering

Electrode Engineering



Figure 4: Three electrodes shown connected to the wires leading to the circuit. From left to right; reference, working, and counter electrode. The working electrode has been extended using silver epoxy.



Figure 5: The working electrode will invade the body and react with intramuscular to create a current. The signal will follow the wire into a circuit where a current will be converted to a glucose concentration. Width not to scale.

Signal Processing

- Three electrode potentiostat utilized for chemical to electrical transduction. 1. Maintains constant + 300 mV (vs. Ag/AgCl reference) at working electrode 2. Converts current produced from redox reaction at working electrode into a voltage - Arduino Uno and Arduino IDE were utilized for data analysis.



Figure 7: Block diagram of circuit. This block diagram outlines how the Arduino, circuit, electrodes, and PC interact with one another to ultimately take the input voltage from the Arduino and turn it into a glucose reading. Arduino IDE is used to make the voltage to glucose conversion.



Figure 8: Three-electrode potentiostat circuit diagram. This circuit diagram depicts the overall configuration of the Arduino controlled potentiostat that was used for testing. There are three main components of this circuit – the voltage divider (1), the control circuit (2), and the transimpedance amplifier (3). The LM324 quad op-amp was the only op-amp used, and it was supplied with +6.5 V and 0.0 V using a DC power supply.

Delivery Model

- Synthetic model to observe electrode delivery and invasiveness
- Stylet guided entry system leads electrodes through various depths
- Sponge mimics depth of body, meat represents the compartment
- Electrode was not connected to circuit and model shown to subjective observation
- Successfully penetrates 8 centimeters



Fabrication Scheme

Signal Processing

Delivery Model



Figure 6: Chloriding the silver wire in a 1.23 M potassium chloride solution. The reference electrode, shown in green, is connected to the cathode of the 9V battery.





Figure 9: Three cross sections shown of electrode invasiveness at 4, 6, 8 centimeters. Models are shown to verify that an electrode can be successfully inserted in a highly invasive way. Insertion ranges vary to accommodate multiple depths ACS can occur within a muscle.

- Glucose Meter



Electrodes

- **3-Electrode Potentiostat**
- amplifier
- stage of the potentiostat

Stylet Insertion Needle

- any compartment

- Calibrate prior to use

Acknowledgements

- Dr. Christopher Doro, Client
- Coordinator
- Consultant





Testing and Results

Initial Probe Testing: Nine PBS/Glucose solutions

- 0, 25, 50, 60, 70, 100, 300, 500, and 1,000 mg/dL glucose concentrations - Concentrations of solutions were measured with the Contour Next ONE

Discussion

Reverse engineered a commercially available working electrode Further development required to obtain a custom built glucose monitor Unknown chemistry of the working electrode lead to difficulties with the interaction between the reference, counter and working electrodes Consumer market glucose sensor is most sensitive in higher ranges and less accurate within our range of interest

Designed and built a potentiostat control circuit and transimpedance

Verified expected voltage differences between electrodes and after each

Incorporated a readily available stylet system to effectively penetrate into

The stylet housed glucose sensor from muscle environment

Future Work

Demonstrate feasibility on liquid and muscle models

Remake electrodes from scratch by coating platinum wire with a more specialized monitor that can accurately read 50 dL/mg to 200 dL/mg

Shrink electrode system into one insertion device

Translate design to be usable in hospital, trauma settings, and active military fields for on-site ACS diagnosis

Professor Walter Block, Advisor Mr. Alexander Siy, Research

Dr. Amit Nimunkar, Bioinstrumentation

References

Hand, 2017, [Online], Available: http://www.assh.org/handcare/hand-arm-iniuries/broken-bones Sitzman, and R. VO, "Can intramuscular glucose levels diagnose compartment

syndrome?," Trauma Acure Care, vol. 76, no. 2, 2014. [4] S. Vaddiraju, D. J. Burgess, I. Tomazos, F. C. Jain, and F. Papadimitrakopoulos, "Technologies for Continuous Glucose Monitoring: Current Problems and Future Promises," J. Diabetes Sci. Technol. J Diabetes Sci Technol, vol. 44, no. 66, pp. 1540–1562, 2010. [5] O. Jones, "Muscles of the Posterior Compartment of the Thigh," *Teach Me Anatomy*, 2014. [Online]. Available: http://teachmeanatomy.info/lower-limb/muscles/thigh/hamstrings/. [Accessed: 06-Dec-2017].