



# ABSTRACT

- Patients with asthma present eosinophils in the lumen of their blood, airways, and inflamed lung tissue
- Eosinophils have different surface markers depending on location in lung
- Project goal: successfully dissociate eosinophils to analyze their surface markers after an asthmatic reaction
- The design should minimize cell damage while dissociating enough cells to run flow cytometry (20,000 cells)

# BACKGROUND

### **Motivation**

• Eosinophil research can lead to advancements in drug delivery mechanisms and further knowledge of asthmatic physiology

### <u>Research</u>

- gentleMACS has been used for over a decade on various rodent lungs, from Guinea pigs to mice<sup>[2]</sup>
- The gentleMACS tube dissociates tissue using various ridges, rotors, and spacers<sup>[2]</sup>
- Why not a microfluidic device?

Reynolds Number =  $\frac{\rho v L}{m}$ 

If L = 0.0001mRe>3000<sup>[4]</sup> to achieve turbulence If  $\rho = 1000 \frac{kg}{m^3}$  and  $\mu = 0.00089 \text{ Pa*sec}$  $v(velocity) = 26.7 \frac{m}{sec} \text{ or } 60 \text{ mph}$ Cells would die instantly upon wall collision

## **DESIGN CRITERIA**

### **<u>Client Requirements</u>**

- Dissociated cells must be viable and have and unchanged surface markers
- Duration of dissociation must be less than 4 hours
- Cost of single use device must be less than \$10 and sterilizable

## <u>Hydrogel Requirements</u>

- Stiffness, swellability, and size adjustability
- Cell should not attach or interact chemically with the hydrogel material
- Beads should not decompose in enzyme solution
- Biocompatible





# Lung Tissue Biopsy Dissociation

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# FINAL DESIGN



*Figure 1: Eosinophil*<sup>[1]</sup>



Figure 2: GentleMACS conical *tube*<sup>[3]</sup>



- 1-2 mm<sup>3</sup> tissue samples were soaked in .15% collagenase and 5 mM Ca for 20 minutes at  $37^{\circ}$ C
- 4.4 g of hydrogels, 2.4 ml enzymes, and 4 pieces of tissue biopsies were added to gentleMACS conical tube
- To dissociate the tissue, the lung tissue setting on the gentleMACS machine was used three times

## **TESTING AND RESULTS**



Figure 4: Number cells alive (black) and total cells (red) in both control and experimental (hydrogel) conditions as counted in the hemocytometer with Trypan Blue



*Figure 6: Comsol simulation with no hydrogel beads.* [Top] There is little pressure against the surface of the tube without beads. [Bottom] The flow of particles (water molecules) is more uniform





Figure 3: Hydrogel Synthesis with beaker of CaCl<sub>2</sub>, stir bar, and pipette of Alginate



Figure 5: Tube with hydrogels and tissue samples before dissociation



Figure 7: Comsol simulation with hydrogel beads. [Top] Greater overall pressure applied to all surfaces [Bottom] Flow of particles more chaotic and "turbulent"

- doesn't degrade in the enzymes
- sample
- to make small enough hydrogels.
- cells
- tissue

- surface markers

# ACKNOWLEDGEMENTS

- Dr. Sameer Mathur
- Dr. Wan-Ju Li
- Paul Fichtinger

- [1] Blausen, B., *Eosinophil Granulocyte*. 2014.

- [Accessed: 02-Dec-2018].
- [3] Miltenyi Biotec, gentleMACS C Tube. 2011.

# DISCUSSION

• Dissociation with enzymes alone in the gentleMACS machine does not result in appreciable dissociation, samples are too small • Comsol simulations (Figure 6 and 7) show hydrogel particles aid in mechanical dissociation by increasing the number and force of interactions the tissue has with surroundings

• Homogenization using steel and glass beads is common; these are too stiff and lyse cells; hydrogels were chosen as replacements • Any biocompatible hydrogel material could be used for dissociation if size and stiffness of beads can be controlled and the hydrogel

• Size of the beads should be approximately the size of the tissue

• The 3.5% sodium alginate was used to create the hydrogel beads; the 2% sodium alginate was not stiff enough and fell apart during the dissociation and a 5% sodium alginate solution was too viscous

• Hydrogels used during final testing cross linked with a 200 mM CaCl<sub>2</sub> dissociated more cells than 100mM CaCl<sub>2</sub>

• If Ca<sup>2+</sup> is required for enzyme activity (i.e. collagenase), alginate is not ideal hydrogel source; alginate expelled into solution is

crosslinked into small particles obscuring analysis and culture of

• A limitation of this design lies in the fact that it requires the gentleMACS dissociator to agitate the hydrogels and dissociate the

## **FUTURE WORK**

• Synthesize and test different hydrogel material

• Conduct additional tests to determine optimal bead size,

swellability, and stiffness for maximum tissue dissociation

• Run final design with tissue sample of inflamed human lung tissue

• Use flow cytometry to determine types of dissociated cells and their

• UW Biomedical Engineering Dept. • Dr. John Puccinelli

# REFERENCES

[2] Jungblut, M., Oeltze, K., Zehnter, I., Hasselmann, D., and Bosio, A., "Preparation of single-cell suspensions from mouse spleen with the gentleMACS Dissociator," J Vis Exp. vol. 11, no. 22, p. 1029, December 2008. [Online]. Available: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2762924/

[4] Schlichting, H., and Gersten, K., Boundary-Layer Theory. Springer Verlag, 2017, p. 416-419.