



# Miniature Microscope for FRET Microscopy

Kaitlyn Gabardi, Kadina Johnston, John Rupel, Ethan Nethery, Benjamin Ratliff  
Client: Professor Matthew Merrins  
Advisor: Professor Jeremy Rogers

## Abstract

Microscopes are used to study the structure and function of cells, and fluorescence microscopy is one method of observing them. In Fluorescence Energy Resonance Transfer (FRET), one can study intracellular signal transduction and molecular interactions. Our client, Professor Matthew Merrins, currently teaches a human biochemistry course at the University of Wisconsin Madison where his students use a laconic biosensor and FRET to study lactate in pancreatic beta islet cells [1]. Currently, he has only one microscope with these capabilities, limiting the enrollment of his course as well as the lab options. These microscopes typically cost over \$100,000; however, they are extremely adaptable. For this course, they will be doing the same lab year after year, so the microscope will only be used for a single purpose. Therefore, the team will design a low-cost microscope for the specific wavelengths of interest.

## Background/Motivation

### FRET – Fluorescent Resonance Energy Transfer:

- Energy transfer between two light-sensitive molecules (chromophores)
- Donor molecule absorbs energy from light source
- Donor is excited and emits lower energy photons
- Energy transferred to acceptor and lower wavelength emitted
- Client uses Laconic FRET Biosensor

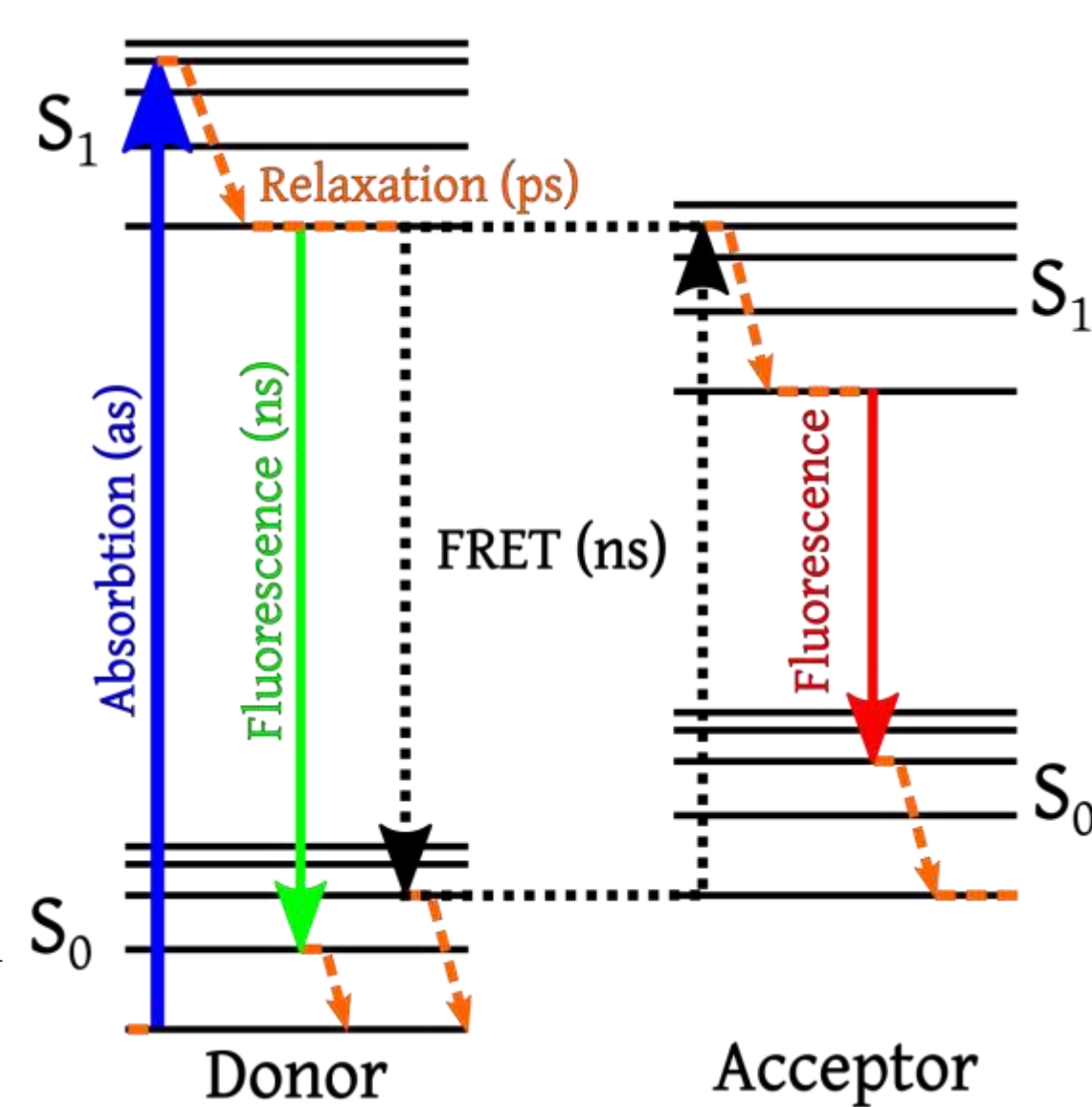


Figure 1. Diagram of FRET concept [2]

### Laconic Biosensor:

- Excitation source of 430 nm
- Donor fluoresces at 470 nm
- Acceptor fluoresces at 535 nm
- Used to measure lactate levels

## Design Criteria

- **Manufacturability:** repeatable for manufacturing with limited previous experience
- **Image Quality:** significant & detectable change in fluorescence between 470 & 535 nm from 430 nm source
- **Cost:** <\$4,000 per microscope
- **Reliability:** consistent results from experimentation, with similar results to client's microscope
- **Operability:** intuitive for student use, easy handling/storage

## Testing

### Comparison of Imaging Algorithms

- Time Series images acquired with Nikon TI Eclipse
- Imaged islets during changes in concentration of glucose
- Measured FRET Ratio over time with professional software
- Measured FRET Ratio over time with amateur software

### Optical Resolution

- Used 200mm tube lens and 0.65NA objective to image test chart
- Calculated resolution from test chart and theoretical resolution

## Results

- Data analysis conducted through a paired T-Test of algorithm's respective FRET ratio slopes over time at  $\alpha = 0.05$
- $H_0: \mu_{Nikon} - \mu_{MATLAB} = 0$
- $H_A: \mu_{Nikon} - \mu_{MATLAB} \neq 0$
- P-Value = 0.71756
- Retain  $H_0$ , suggesting the MATLAB algorithm detected the same relative changes in FRET ratio

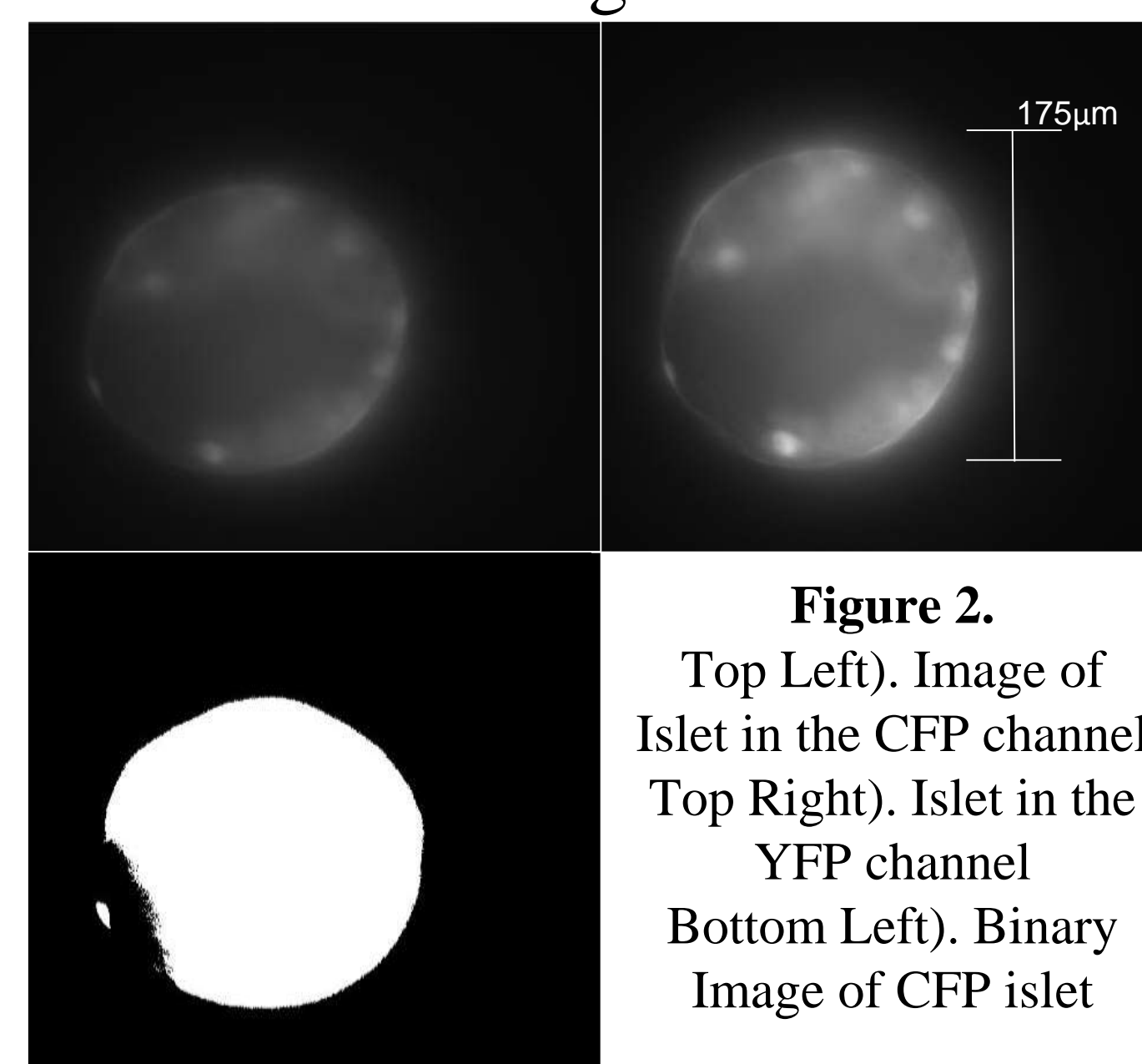


Figure 2. Top Left). Image of Islet in the CFP channel  
Top Right). Islet in the YFP channel  
Bottom Left). Binary Image of CFP islet

- 1951 USAF resolution test chart

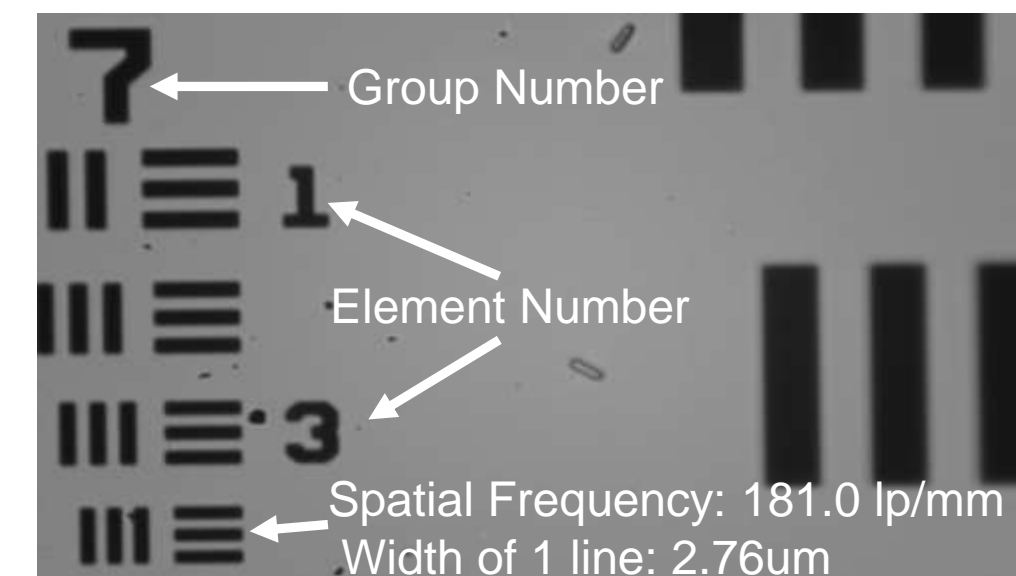


Figure 4. Image of test chart using Imaging Source Camera

### Calculation for Resolution

$$\text{Resolution } (\mu\text{m}) = \frac{1000}{2^{\text{Group} + \frac{\text{element}-1}{6}}}$$

$$5.5\mu\text{m} = (2^{7+(4-1)/6})^{-1} * 1000$$

$$\text{Theoretical Resolution } (\mu\text{m}) = \frac{0.61\lambda}{NA}$$

$$0.511\mu\text{m} = \frac{0.61 * 0.545\mu\text{m}}{0.65}$$

## Acknowledgements

The team would like to extend their gratitude and thanks to Professor Rogers, Professor Merrins, Sophie Sdao, Zach Simmons, Ryan Niemeier, Michael Zupan, MS Paint, BME Department, and the MakerSpace

## Final Design

### Optical Component

- Tube Lens
- Objective
- Filters

### Mechanical Component

- 3D printed parts
- Fold Mirror
- Focuser

### Software

- Control of electro-mechanical components
- Image Processing
- Display

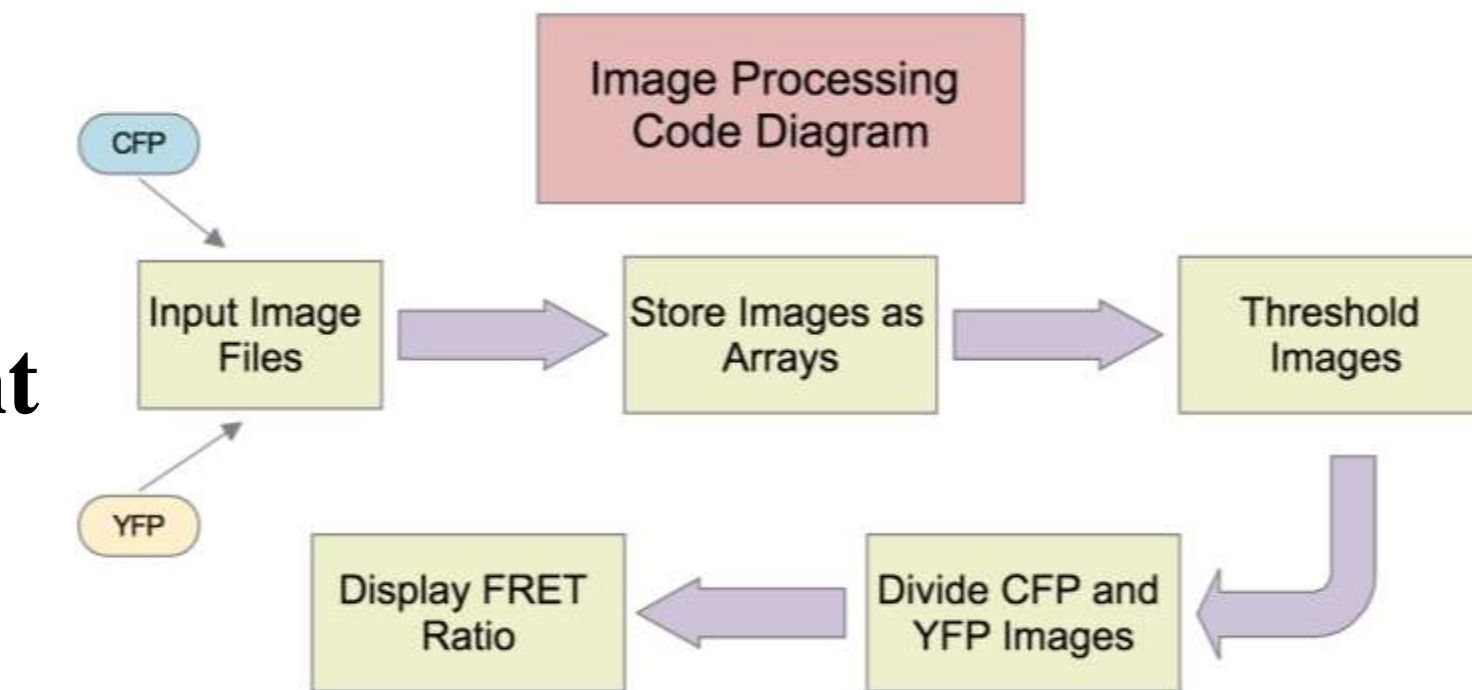


Figure 5. Block diagram covering general protocol of image analysis in software

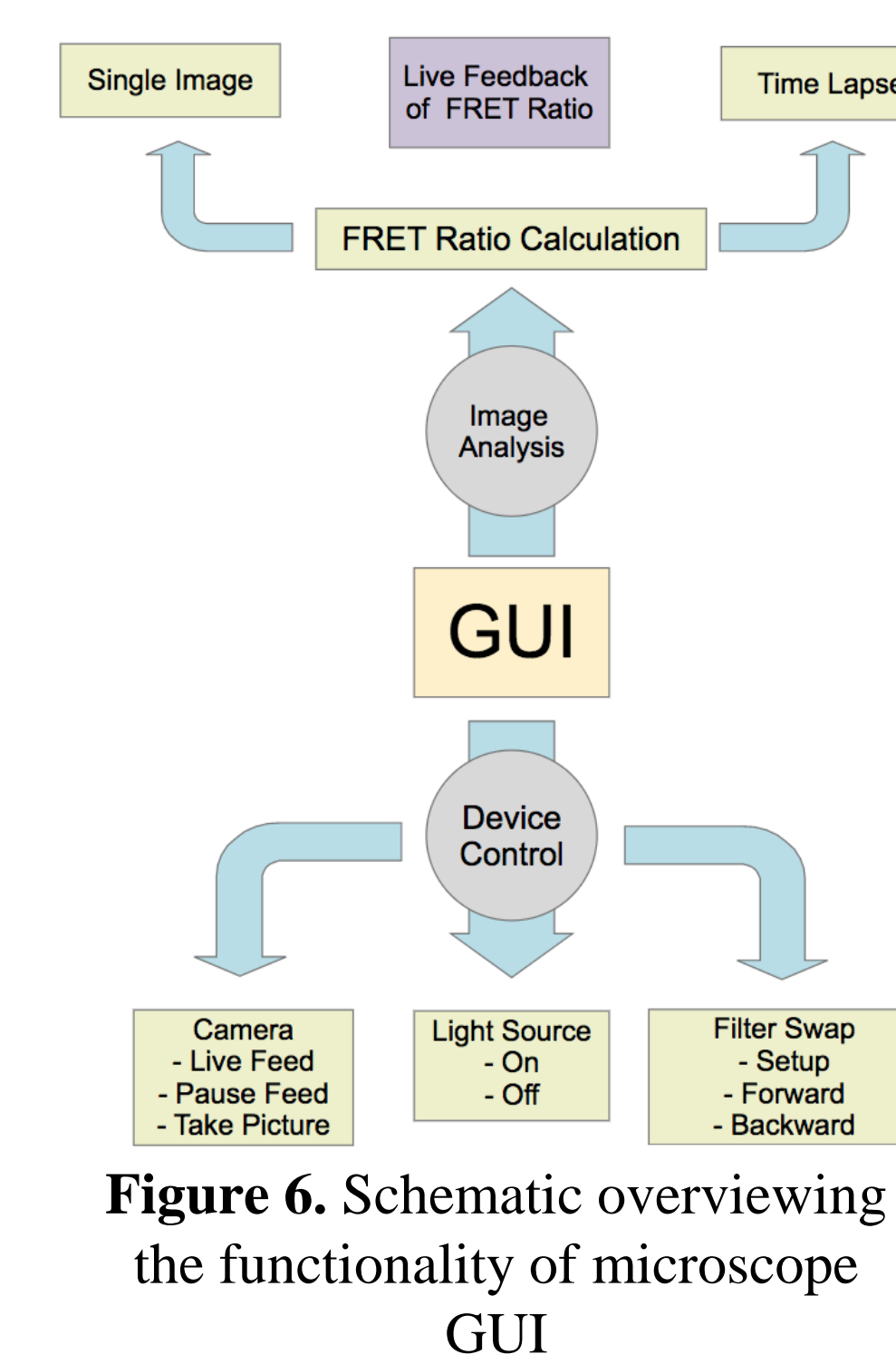


Figure 6. Schematic overviewing the functionality of microscope GUI

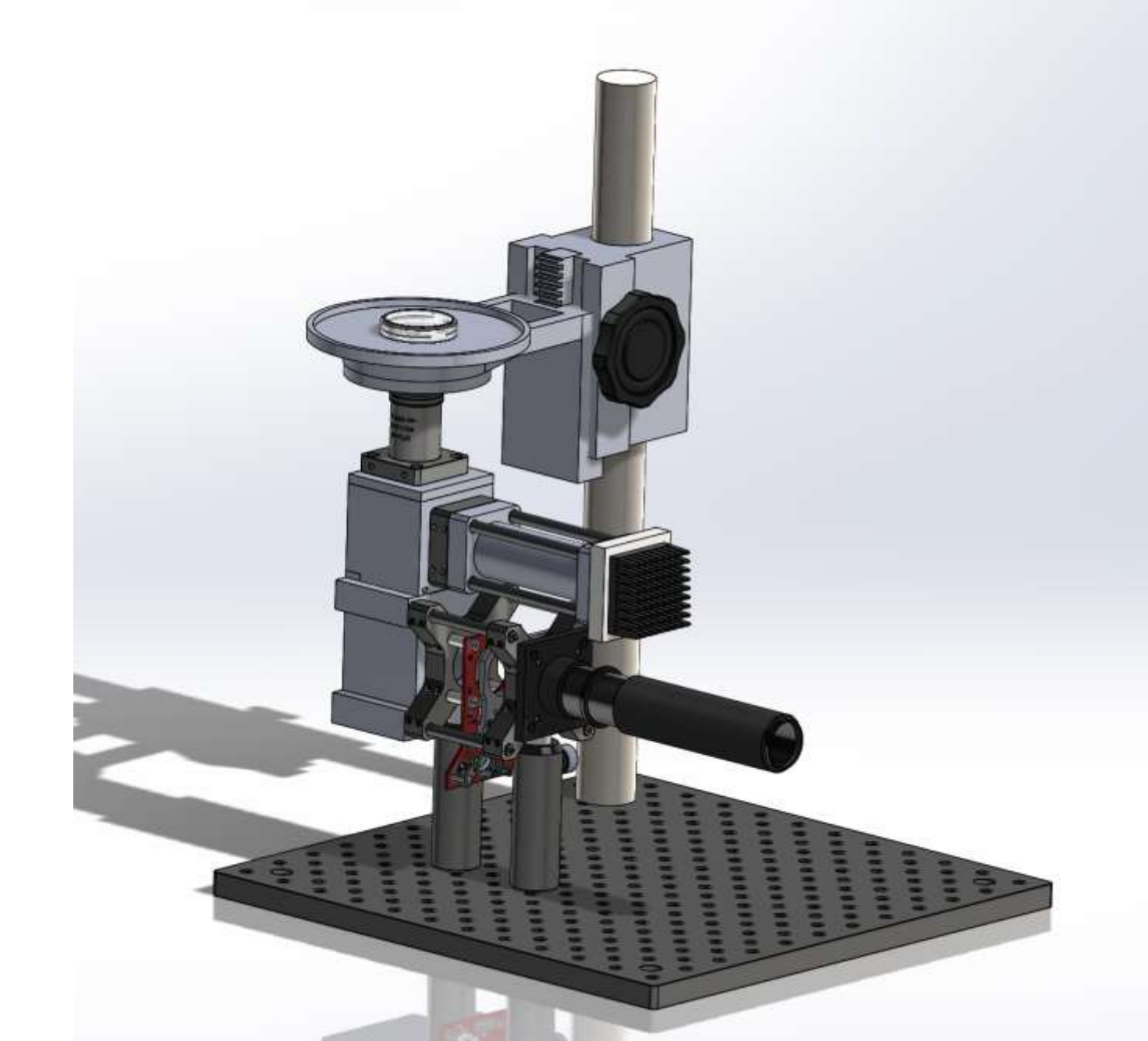


Figure 7. Computer designed rendering of microscope assembly

## Future Work

- Refine the GUI and improve its ergonomics
- Make the LED light source more user-friendly and improve its ergonomics
- Replace the circuitry with a PCB board
- Perform a cost analysis to lower the prototype cost below \$4,000
- Consider ways to improve factors that affect the microscope's image quality
- Improve the accuracy of the FRET calculations and image processing

## References

- [1] San Martín A, Ceballos S, Ruminot I, Lerchundi R, Frommer WB, et al. (2013) A Genetically Encoded FRET Lactate Sensor and Its Use To Detect the Warburg Effect in Single Cancer Cells. PLOS ONE 8(2): e57712. doi: 10.1371/journal.pone.0057712  
[2] Alex M Mooney (https://commons.wikimedia.org/wiki/File:FRET\_Jabolinski\_Diagram.svg), "FRET Jabolinski Diagram", https://creativecommons.org/licenses/by-sa/3.0/legalcode