

Tissue Model of the Epithelial Mesenchymal Trophic Unit

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PROBLEM STATEMENT

- Lack of scaffolds that model the lung ECM and its changes due to cell injury from diseases (like COPD & pulmonary fibrosis)
- Dr. Brasier of the UWSMPH needs such a scaffold
 - Provide him with a tool to study lung inflammation and disease
 - Would aid in translational research such as therapies that target lung tissue changes induced by diseased state
 - Scaffold must be bioprinted

BACKGROUND

- The extracellular matrix (ECM) is a network of proteins and macromolecules [1]
 - Provides support and mechanical/biochemical cues to cells
- The epithelial mesenchymal trophic unit (EMTU) is made of [1]:
 - Lung epithelial cells, surrounding ECM, subepithelial fibroblasts
- Chronic lung diseases injure lung epithelium [2]
 - Inflammatory response increases fibroblast activity
 - Fibroblasts produce more proteins such as collagen and fibronectin
 - The mechanical stiffness of the ECM increases

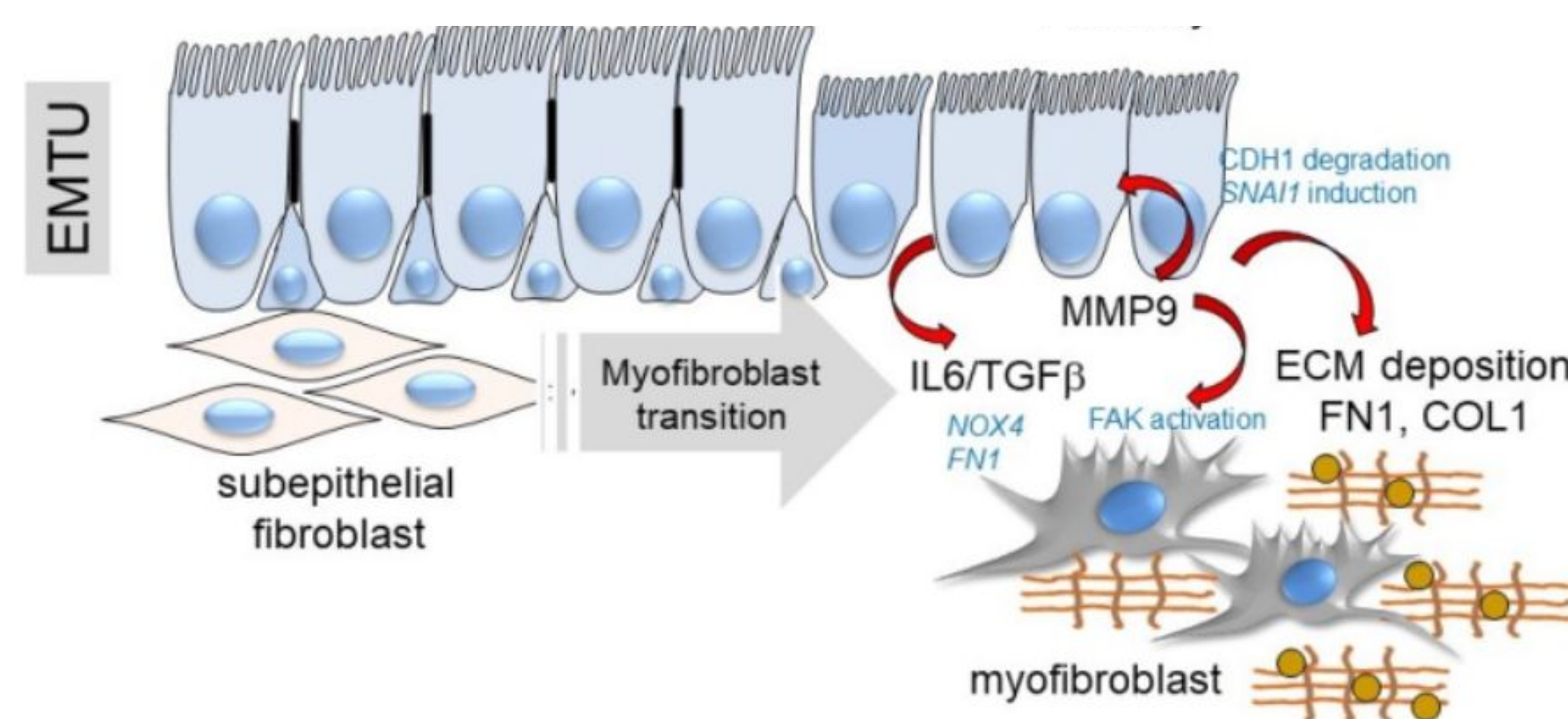


Figure 1: Schematic of EMTU response to injury [2]

MOTIVATION

- COPD is 3rd leading cause of death in the world [3]
- In the US, over 120,000 people die yearly of COPD [4]
- While not curable, current treatments include:
 - Bronchodilators (inhalers) which relax and open the airways
 - Oral steroid medications
 - Pulmonary rehabilitation
 - Surgery (severe cases) [3]
- Tissue model would contribute to more thorough understanding of diseases and development of individualized treatments

DESIGN CRITERIA

- Tunable mechanical stiffness
 - Normal tissue: 2-5 kPa
 - Fibrotic tissue: ≥ 16.5 kPa
- Mimics biochemical properties of native lung ECM
 - Cell adhesive
 - Enzymatically degradable by matrix metalloproteinases (MMPs)
- < 12 mm diameter to be compatible with an air-liquid interface (ALI)

FINAL DESIGN

- **Biomaterial:** Gelatin Methacryloyl (GelMA)
 - **Pipette-based hydrogels**
 - GelMA 50% degree of functionalization
 - Cylindrical molds
 - 9 mm diameter
 - ALI compatible
 - Prior to photocrosslinking gels were allowed to cool/set at 4 °C
 - 365 nm UV light
 - Prior semester stiffnesses
 - Low stiffness: 4.08 +/- 0.56 kPa
 - High stiffness: 24.2 +/- 9.2 kPa
 - **Bioprinted hydrogels**
 - CELLINK GelMA [6]
 - Degree of functionalization not specified
 - LAP at 0.25% incorporated in cartridge
 - 3D cylindrical structure
 - 405 nm UV light

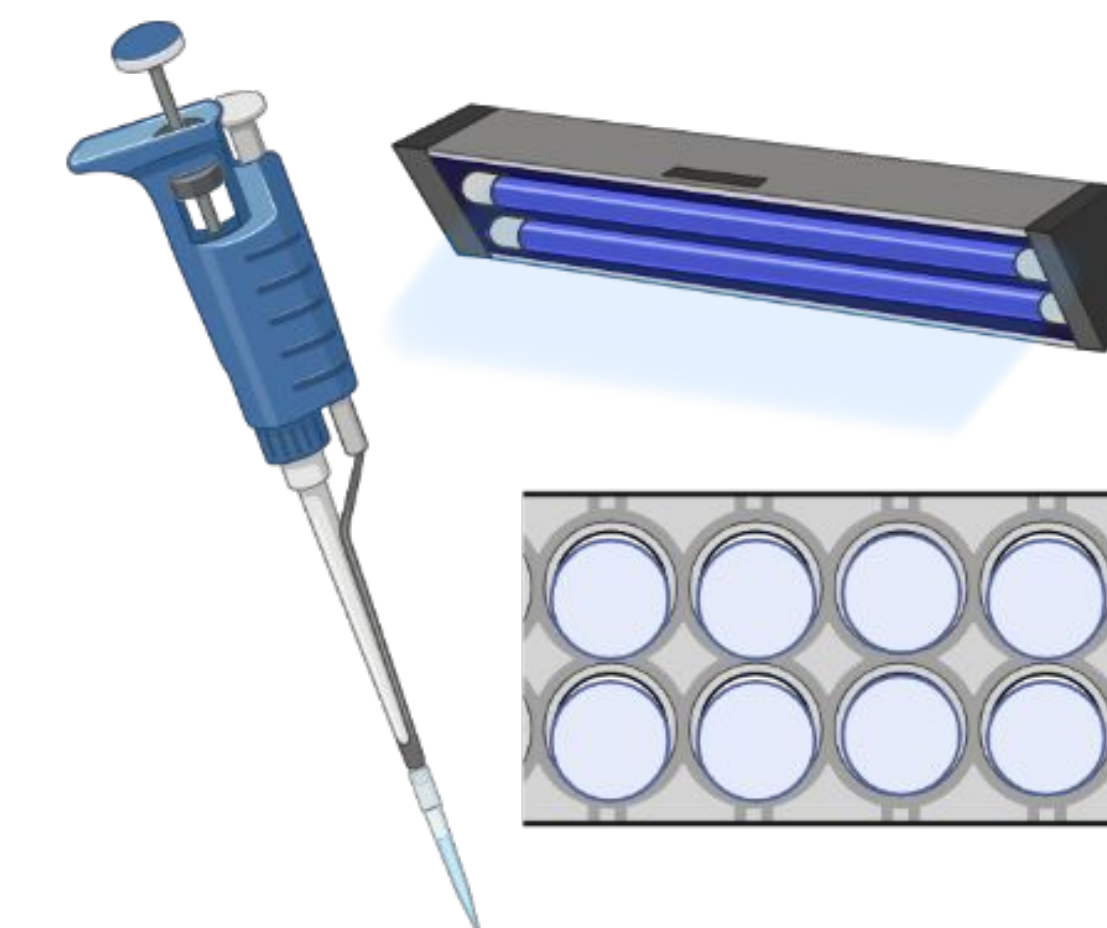


Figure 2: Pipette-based hydrogel tools (made with Biorender)

Figure 4: BioX CELLINK bioprinter [5]

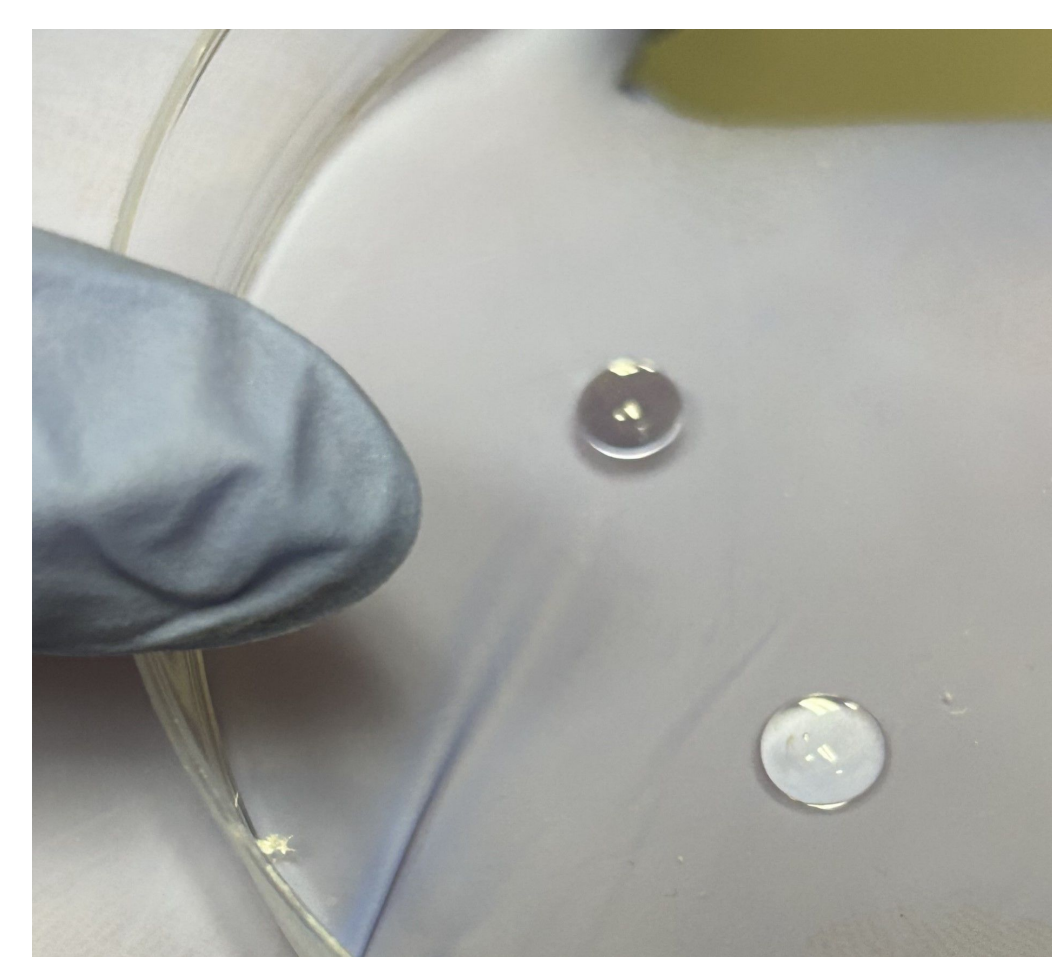


Figure 3: Bioprinted GelMA hydrogels



TESTING AND RESULTS

- **Pipette-based hydrogels**
 - Seeded at 8 million cells / mL
 - Preliminary cell viability test: 81% @ 24 hr
 - Visibly verified cell adhesion and morphology
 - Rheometry (at room temperature)
- **Bioprinted hydrogels**
 - Printer Optimization
 - Increasing equilibrium time and using new GelMA bioink improved printability
 - Heating Temperature and Equilibrium Time
 - Initial: 37 °C and 10 min
 - Final: 37 °C and 20 min
 - Infill density: 35%
 - Printing Temperature
 - Initial: 27 °C
 - Final: 28 °C
 - Extrusion Pressure
 - Initial: 35 kPa
 - Final: 22 kPa
 - New GelMA Stiffness: 995 ± 915 Pa
 - Rheometry (at 37 °C)
 - UV light 2 sec

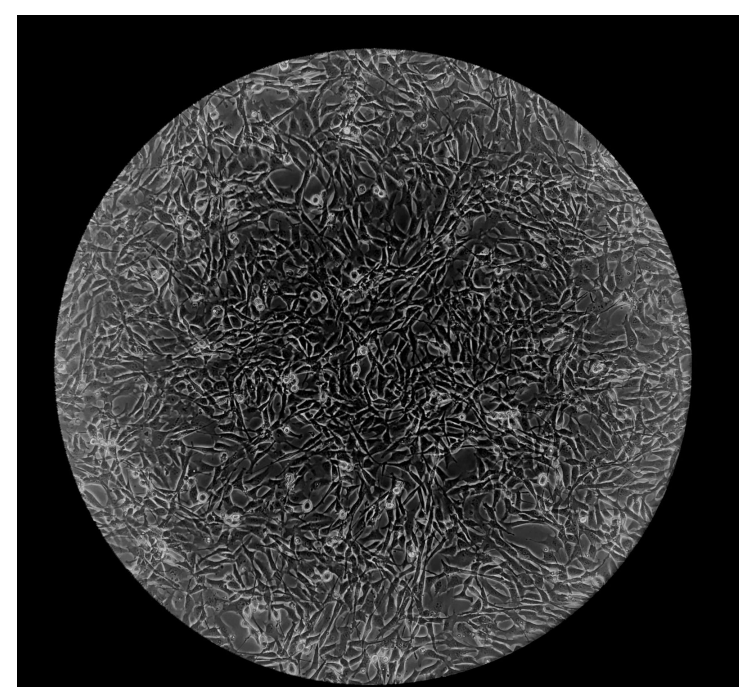


Figure 5: Bright field image after 24 hours in cell culture

Pipette-Based Hydrogels	
Condition (4 °C, UV)	Stiffness (kPa)
3 min, 5 min	4.2 ± 0.92
5 min, 5 min	6.63 ± 2.6
10 min, 5 min	13.24 ± 2.8
5 min, 7 min	49.65 ± 22
5 min, 10 min	277 ± 155

Figure 6: Gels were set in 4 °C prior to UV crosslinking

FUTURE WORK

- Identify and utilize new quantitative cell viability assay
- Longer experimental timescale - matrix remodeling
- Bioprinter optimization
 - Utilize new GelMA bioink
 - Use CAD file to 3D print cylindrical structure
 - Tune UV exposure for fibrotic and healthy ECM stiffness
 - Plan for fibroblast encapsulation and hSAEC monolayer
 - 1×10^6 cells/mL
- Fabricate own GelMA if needed
 - Complete methacrylation, dialysis, and lyophilization process

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