

PROBLEM STATEMENT

- Cancer impacts millions of lives each year, with an estimated 2.2 million new cases and 736,790 deaths reported in the United States in 2024 [1]
- The team was tasked with developing a 3D spheroid formation protocol compatible with genome-wide CRISPR interference (CRISPRi) screening
- Next semester, the team will develop γ H2AX staining protocol to identify sources of DNA double-strand breaks (DSB)
- Following γ H2AX staining, a genome-wide CRISPRi screen will identify factors involved in genome stability

BACKGROUND

- 2D monolayer models have helped study cancer proliferation and therapy responses [2]
- 2D models do not accurately replicate the 3D tumor environment [2]

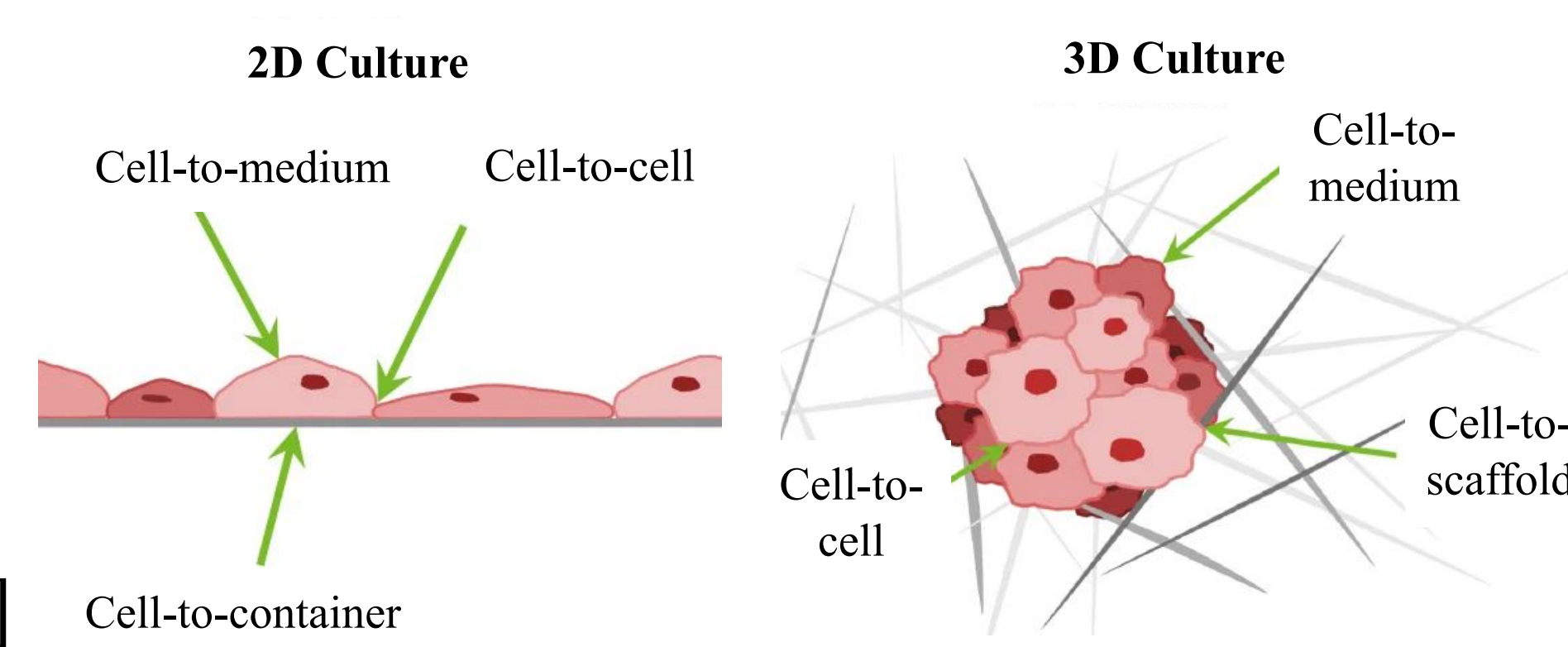


Figure 1: 2D vs 3D culture [3]

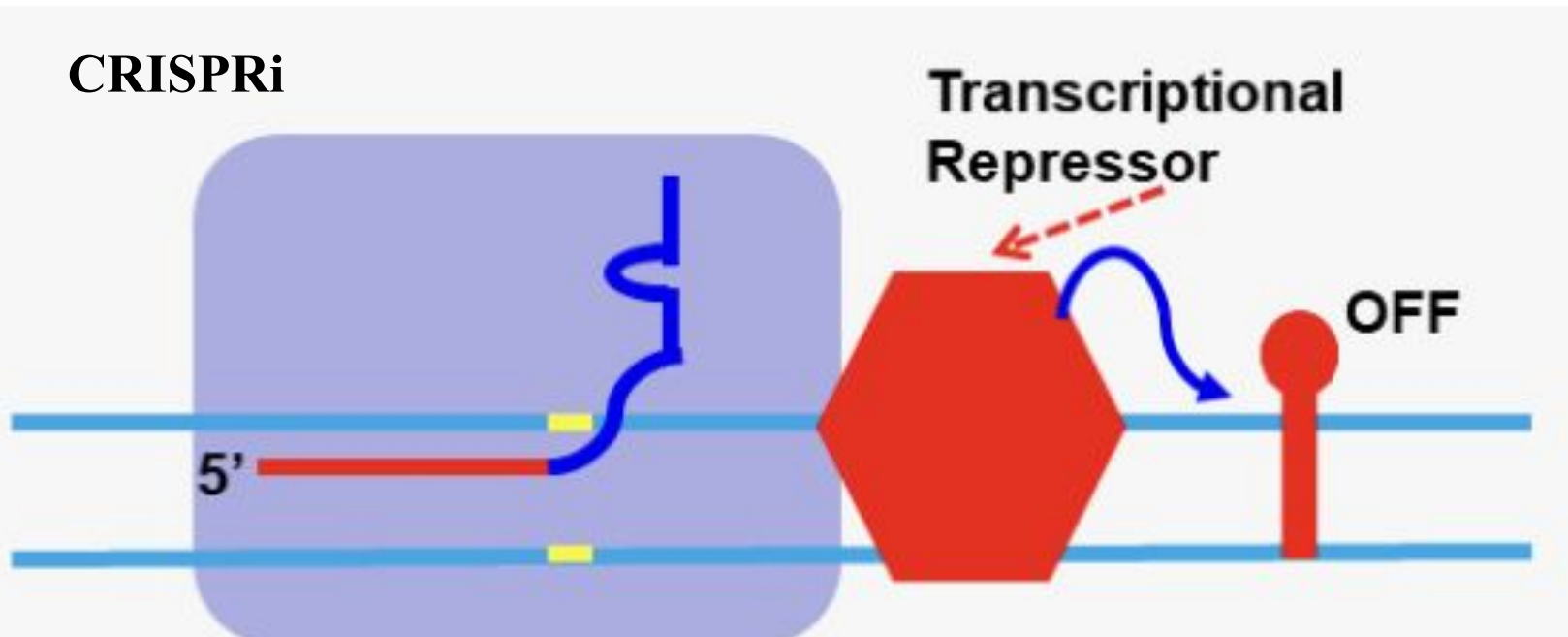


Figure 2: CRISPRi gene knockdown via transcriptional repressor [5]

- CRISPRi via lentivirus allows precise gene repression using deactivated Cas9 (dCas9) without cutting the DNA [4]
- γ H2AX initiates a signaling cascade that recruits DNA repair proteins and facilitates genomic stability [7]

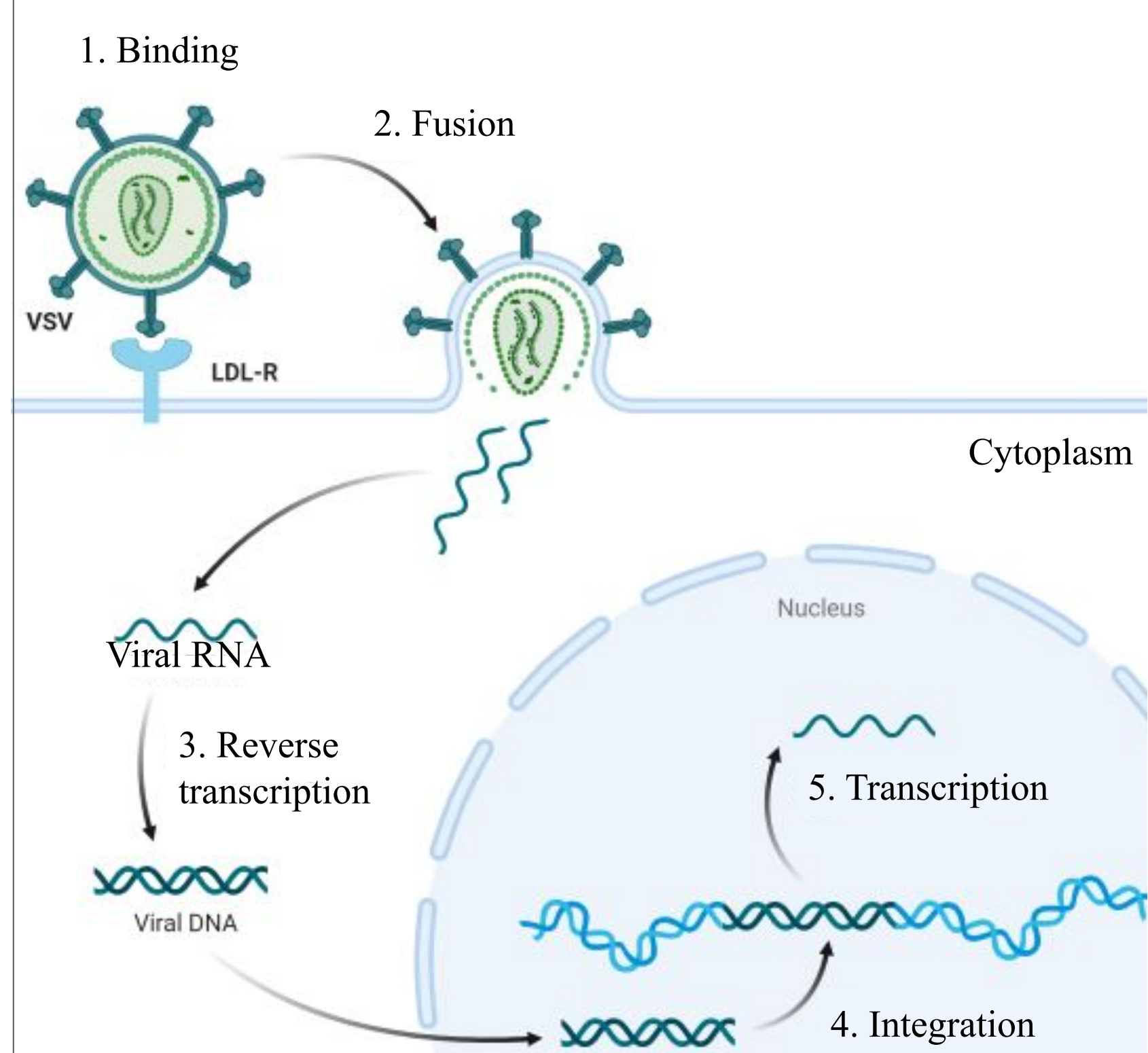


Figure 3: Gene manipulation via lentiviral infection [6]

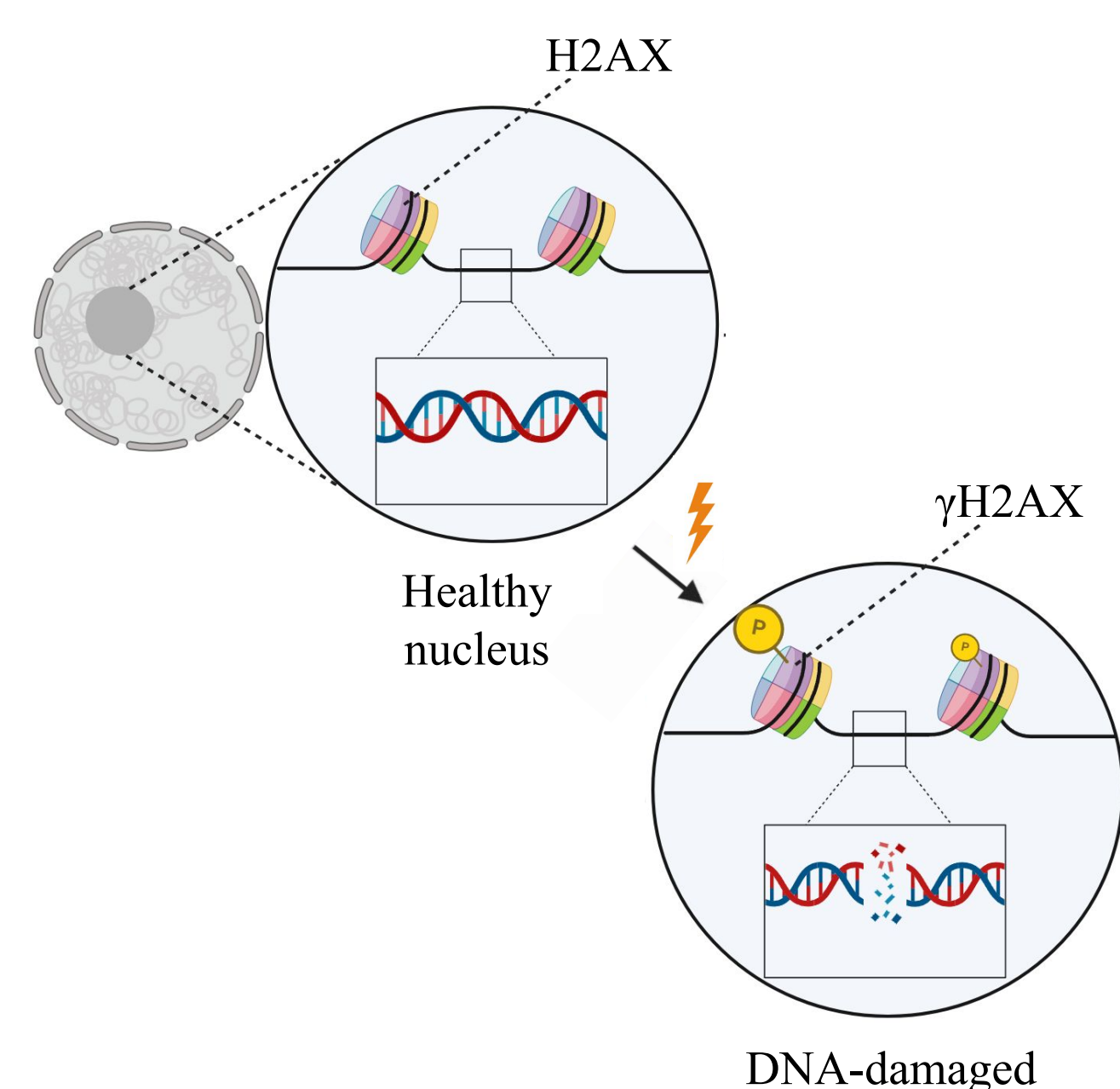


Figure 4: H2AX phosphorylation into γ H2AX during DNA breaks [8]

DESIGN CRITERIA

- ✓ Select suitable cancer cell line
- ✓ Select and optimize spheroid formation protocol
- ✓ Scale 3D culture to prepare for genome-wide CRISPRi screen
- ✓ Adhere to all Biosafety Level 2 (BSL-2) standards
- ✓ Budget of \$1000

FINAL DESIGN

Cell Line: A549

- Non-small cell lung cancer (NSCLC) [9]
 - Adenocarcinoma
 - Adherent
 - 50 μ m cell diameter
- Maintenance
 - Doubling time: 22 hours
 - Confluency 5,000,000 cells/10 mL
- Cells provided by Hess Lab

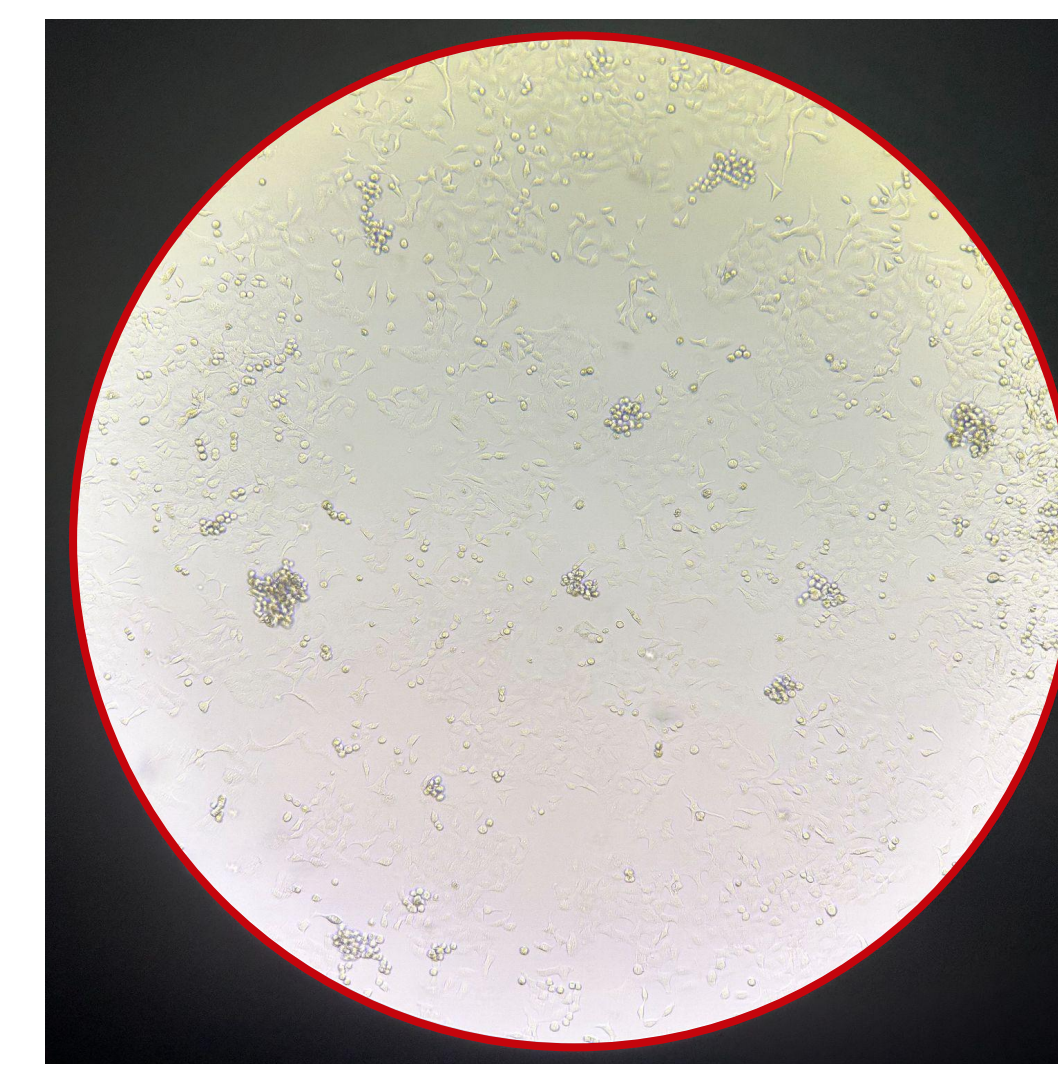


Figure 5: 10x Brightfield image of A549 Passage 5 cells taken at 200% confluency.

PolyHEMA-coated Plates

- PolyHEMA stock solution
 - 1.3 g poly-HEMA
 - 33 mL 99% ethanol
- Add 50 μ L of PolyHEMA stock to each well in a 96-well flat-bottom plates, left dry overnight

Spheroid Formation

- In each polyHEMA-coated well, seed cells in 320 μ L of serum-free DMEM with 0.75% methylcellulose/cm²
 - 4 densities: 25k, 50k, 75k, and 150k cells/cm²
 - 6 wells/seeding density \rightarrow 24 total
- Spheroids will form after 3-4 days
- Spheroids can be dissociated with Accutase (150 μ L/well) [10]

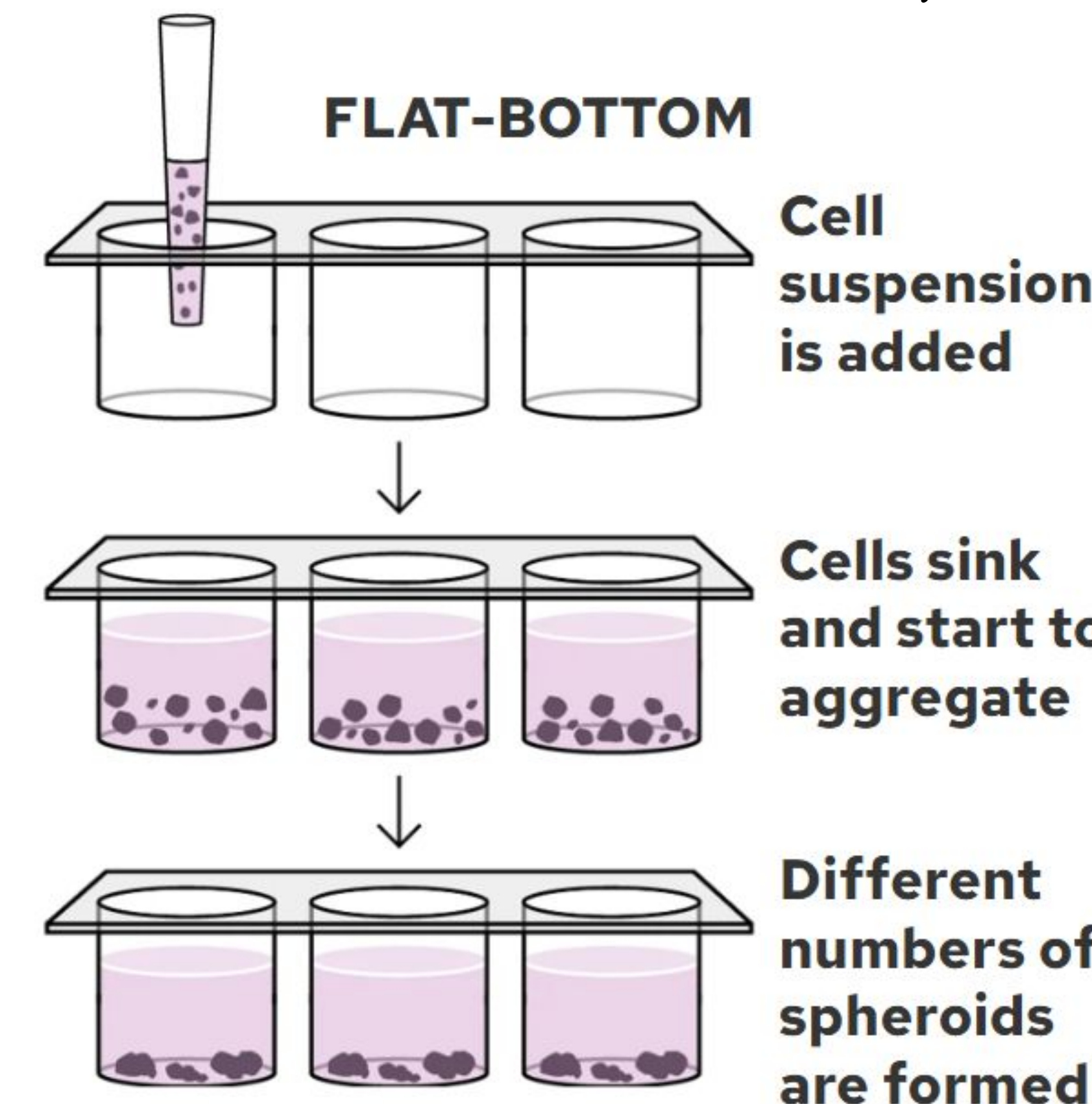


Figure 6: Process of spheroid formation after cell seeding in flat-bottom wells. Picture adapted from Ashgar *et al.*, 2013 [11].

IMAGING

- Used BioTek Cytation microscope to image wells individually
- 5 layers/well \rightarrow 120 total images

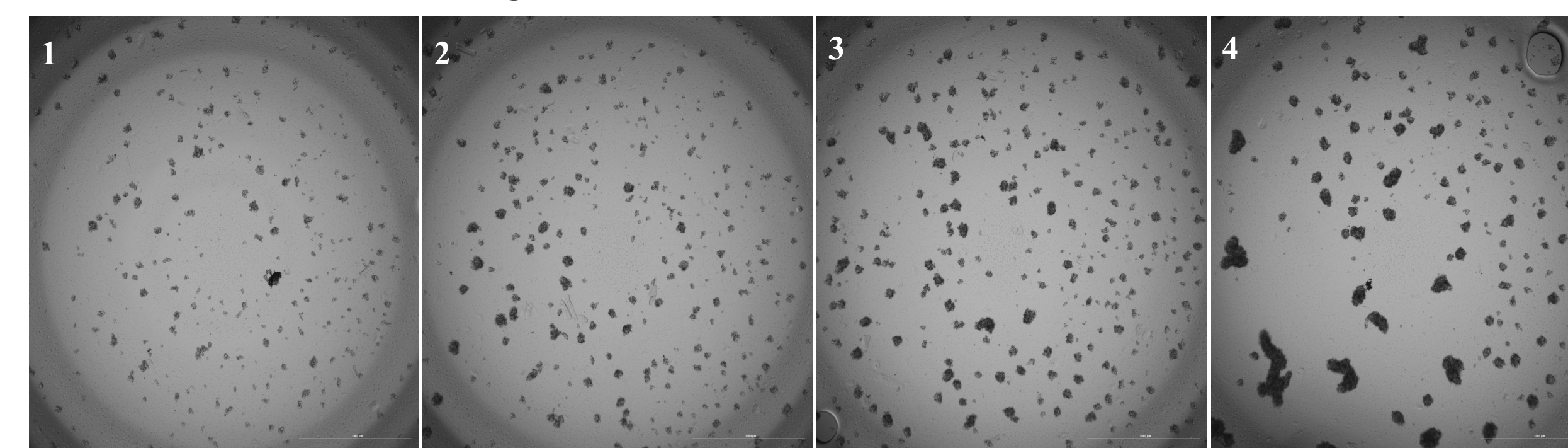


Figure 7: Representative brightfield images of wells 3 days after initial cell seeding for densities 1-4 (25k, 50k, 75k, and 150k cells/cm², respectively).

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ANALYSIS AND RESULTS

ImageJ Analysis

- Used most in-focus images (of 5 layers) and changed to 16 bit
- Applied threshold feature to identify pixels representing cells
- Applied watershed feature to separate clumped spheroids
- Analyzed spheroids with ≥ 20 cells ($\sim 4147 \mu\text{m}^2$) [12]

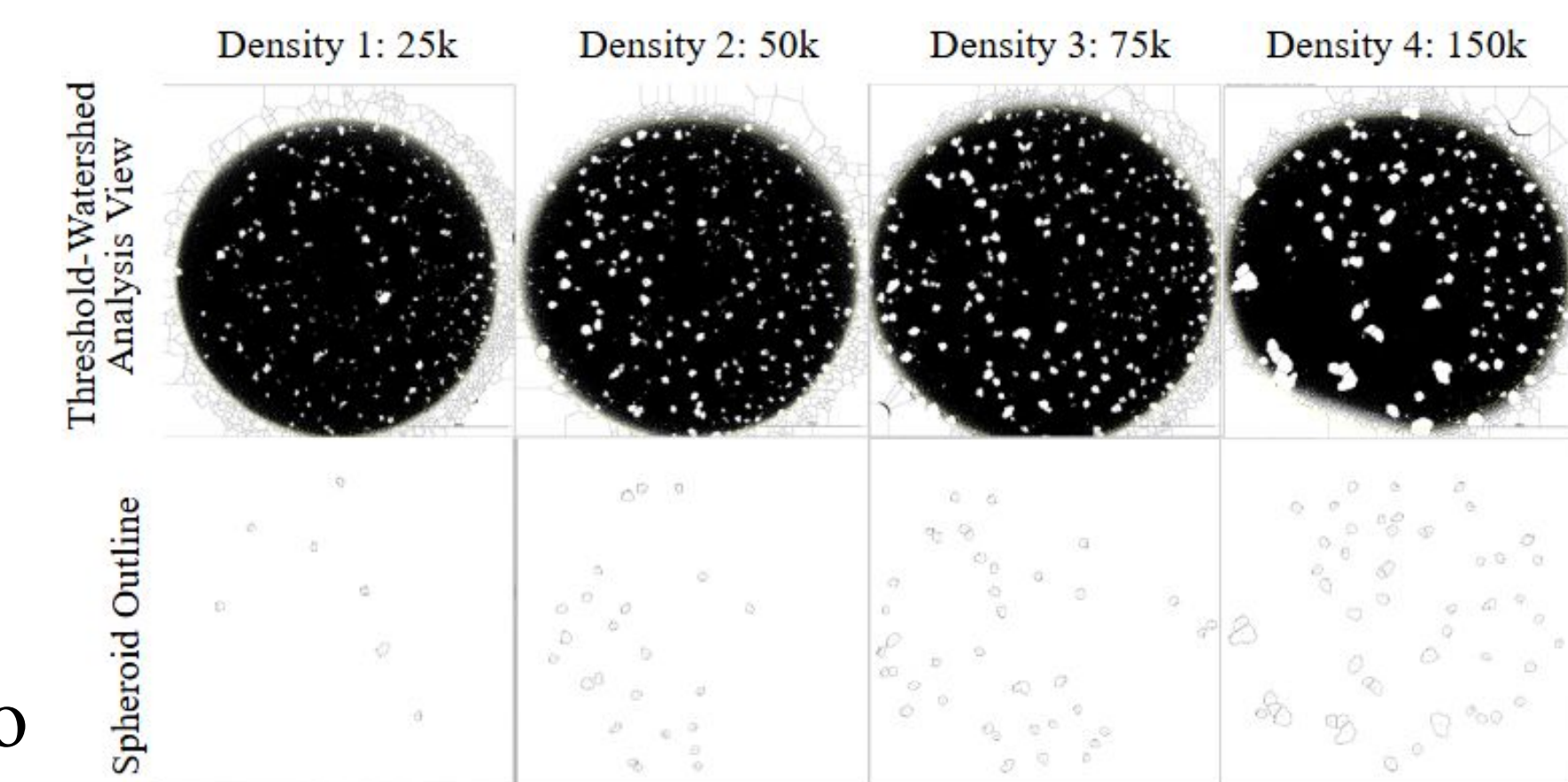


Figure 8: ImageJ analysis of spheroids across seeding densities 1-4 (25k, 50k, 75k, and 150k cells/cm², respectively). Images in top row are in threshold-watershed view and images in bottom row are in spheroid outline view.

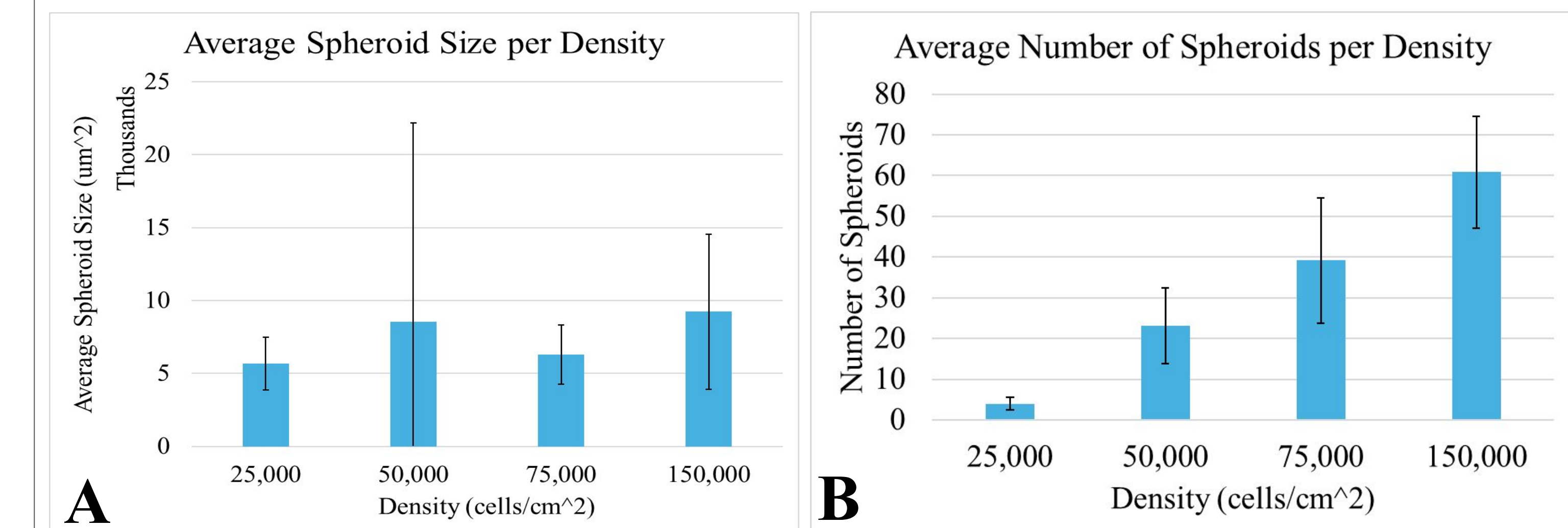


Figure 9: Spheroid analysis across seeding densities 1-4 (25k, 50k, 75k, and 150k cells/cm²). A: Average spheroid size (in μm^2) across seeding densities (in cells/cm²). B: Average number of spheroids across seeding densities (in cells/cm²). Error bars in A and B display standard deviation.

DISCUSSION & FUTURE WORK

Discussion

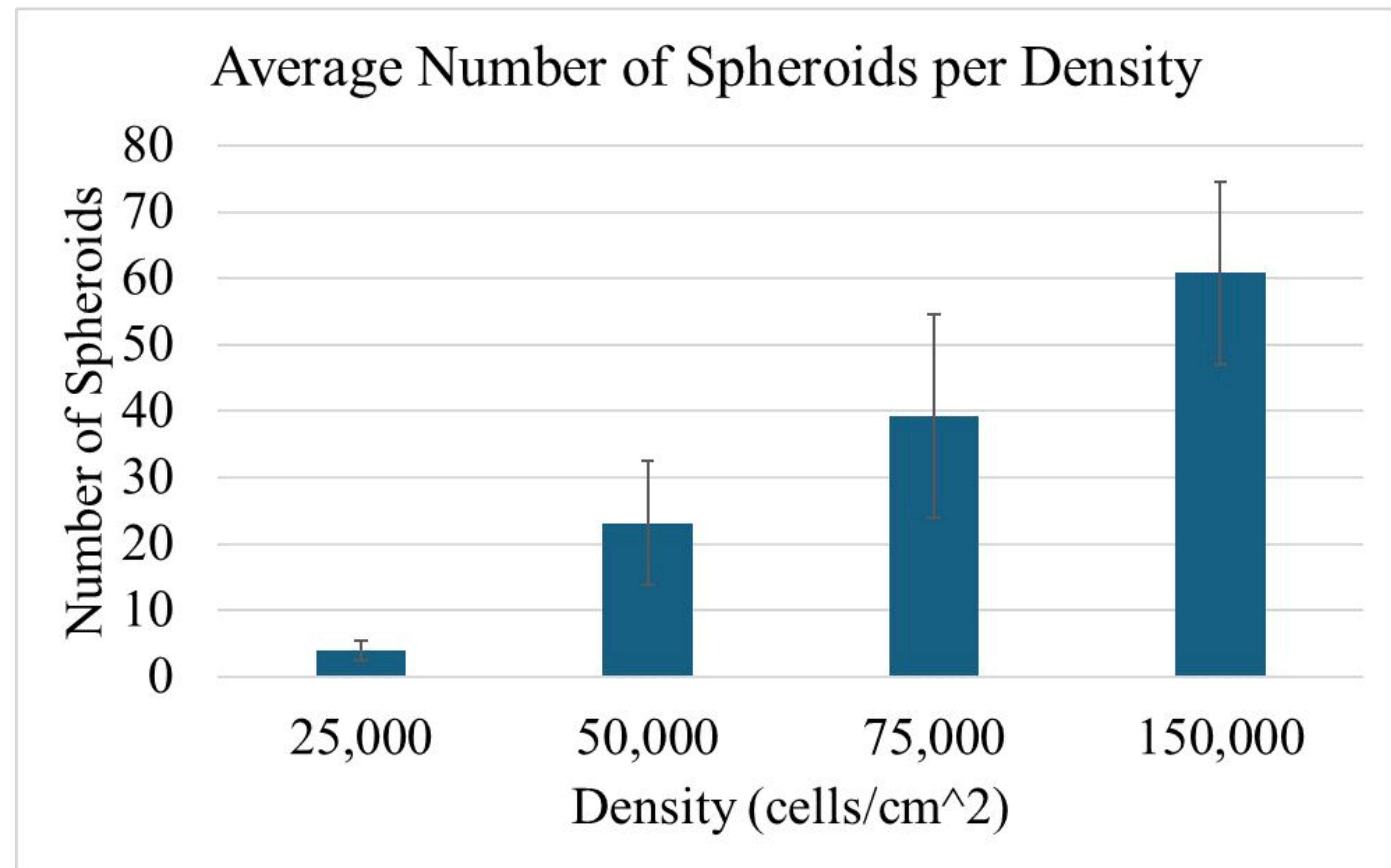
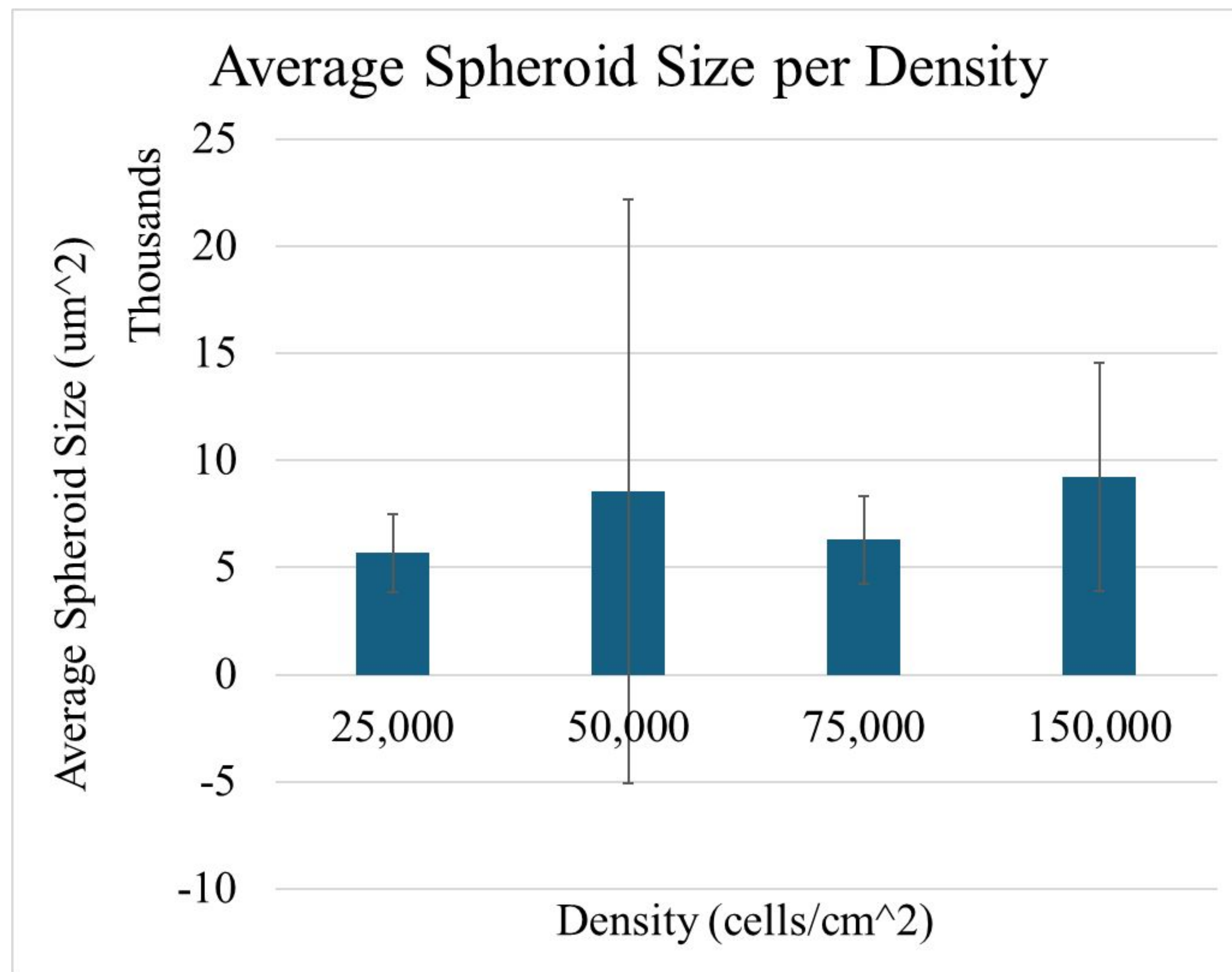
- Cultured viable, adherent human cell line A549 under BSL-2 guidelines
- Established passaging protocol reflective of A549 doubling time
- Average spheroid size remained steady and average spheroid number increased linearly with seeding density
- Largest average spheroid size was within the 50-800 μ m diameter range typically observed in literature [13]

Future Work

- Repeat trial of seeding density variation with % live/dead cytometry assay to confirm preliminary results
- Optimize methylcellulose concentration for spheroid formation
- Perform qPCR for SOX2 to confirm gene expression changes for 3D (spheroids) vs. 2D [14]
- Optimize dissociation step in γ H2AX staining protocol

ACKNOWLEDGMENTS

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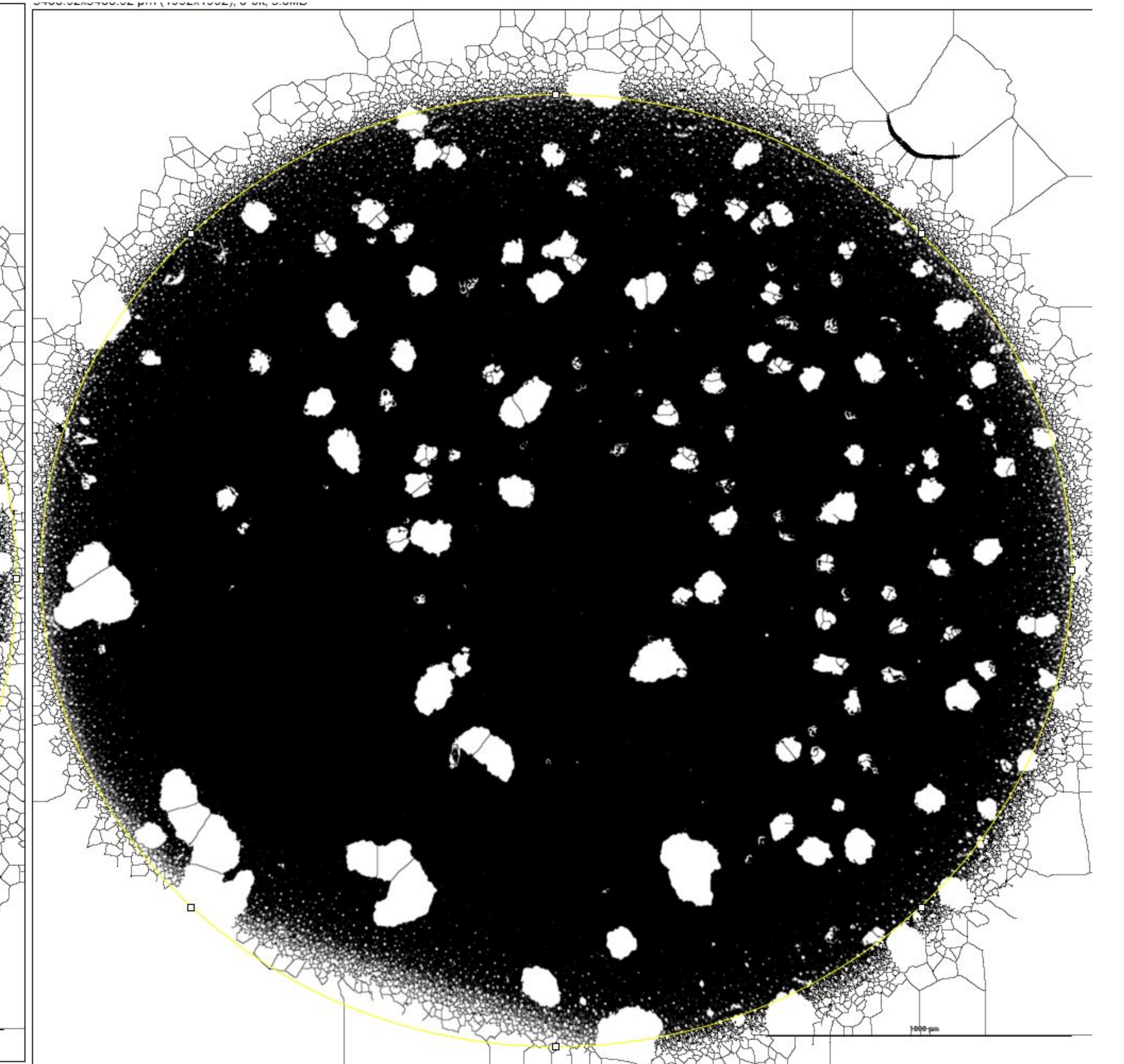
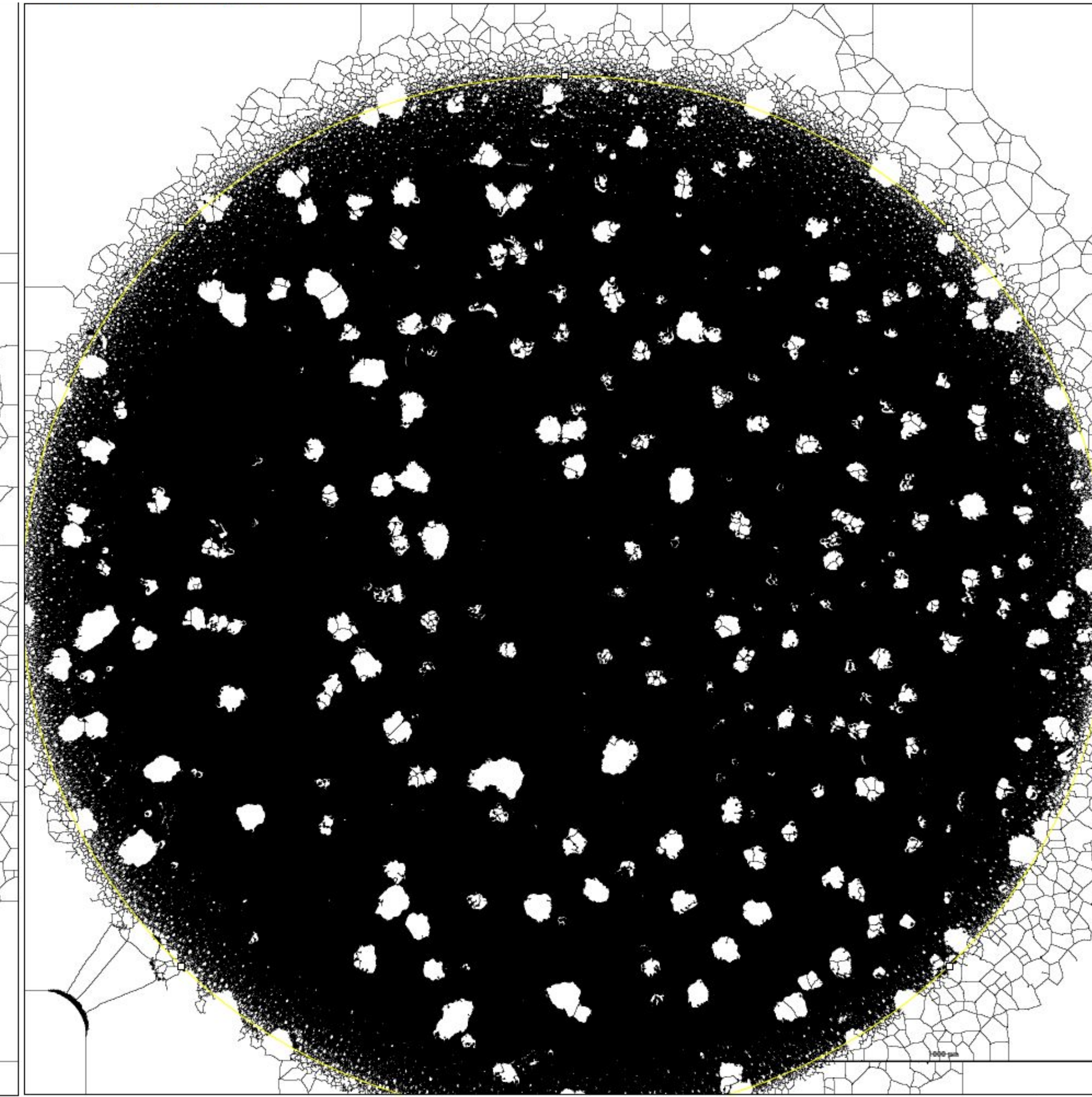
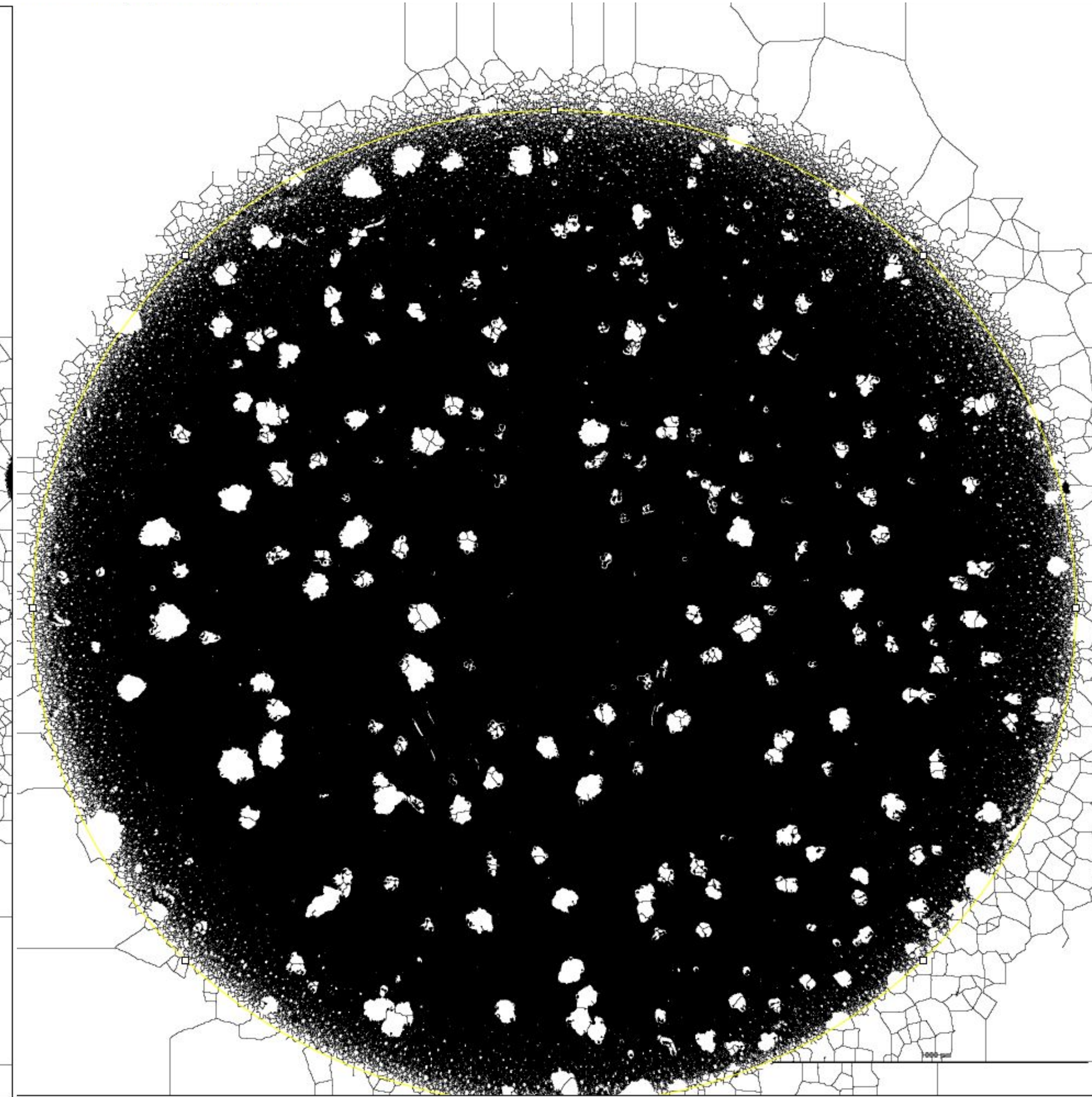
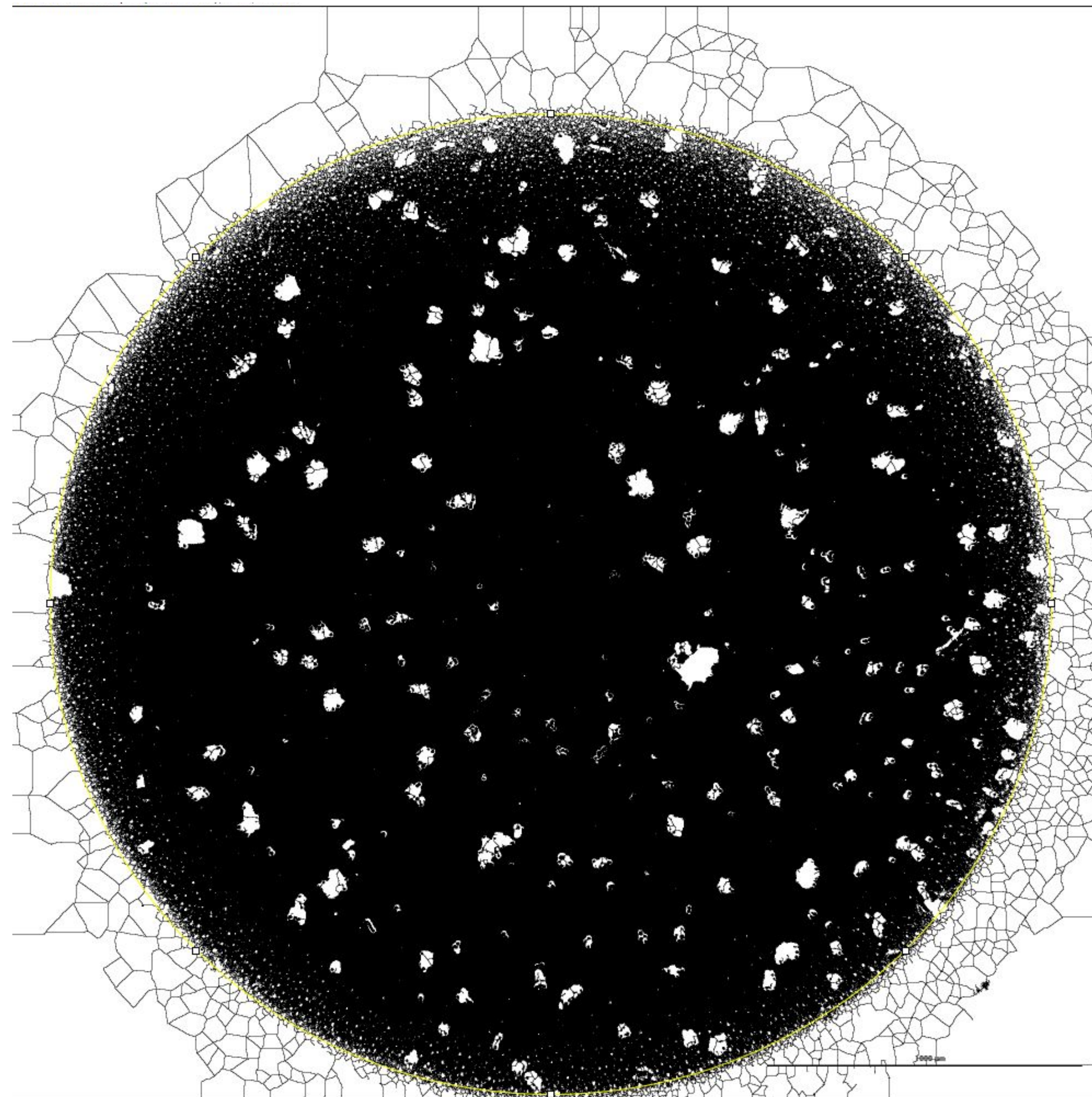
Threshold-Watershed
Analysis View

Density 1: 25k

Density 2: 50k

Density 3: 75k

Density 4: 150k



Spheroid Outline

