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## ABSTRACT

Live cell imaging experiments are difficult to perform over long periods of time on normal lab microscopes. The client desires an inexpensive on-stage incubation chamber that is capable of maintaining temperature, CO<sub>2</sub>, and humidity evenly throughout the chamber at a physiological set point. An initial prototype has been developed that involves a small, cohesive system to regulate these parameters through a feedback systems. Further development of the design will further test and refine the hardware and feedback systems, ultimately bridging the gap in the market between high-cost, functional systems and cheaper, less effective systems.

## BACKGROUND

- Real time imaging offers researchers the opportunity to perform new assays
- Current market in need of affordable and more robust system for real time imaging in cell culture
- Mimicking the physiological environment requires control of temperature, humidity and CO<sub>2</sub> concentration
- Optical compatibility: desired magnification (focal length) and size limitations

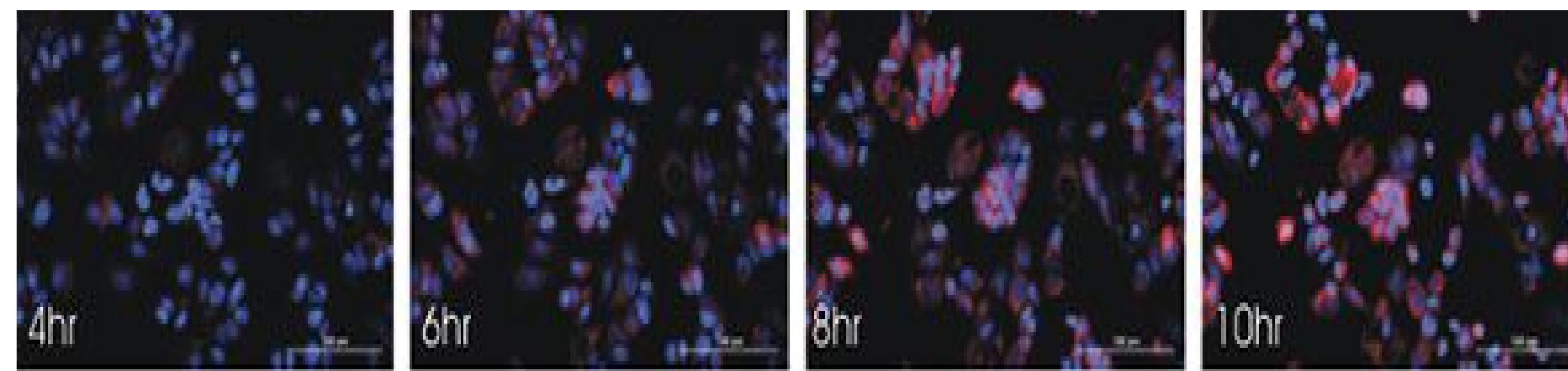


Figure 1. Sample time-lapse imaging using cell fluorescence.

## DESIGN CRITERIA

- Environmental Controls for Physiological Maintenance:
  - Temperature: 37°C ± 1°C
  - Relative Humidity: 95% ± 5%
  - CO<sub>2</sub>: 5% ± 0.5%
- Recovery: Temperature and CO<sub>2</sub> recovery in 6 seconds after 30 second chamber opening, comparable to current products
- Compatible with various microscopes

## MARKET DEMAND

- Limitations of current market
  - High end systems
    - Expensive & limited to one microscope
  - Low end systems
    - Offer poor environmental control
- Our Design Attributes
  - Simplicity
  - Affordability



Figure 2: Low end system (4).



Figure 3: High end system (5).

## DESIGN SOLUTION

- Control systems independently validated
  - integrated to regulate CO<sub>2</sub>, RH, and temperature effectively
  - LCD display of environmental conditions
- 3/32" glass below cell culture to optimize imaging
- Removable lid for fast plate removal

### Design Costs

Design Area	Product names	Category Cost
Environment Sensors	DHT-22, MHZ -16	\$80.05
Environment Control Components	Fluid valve, Immersion Heater	\$50
Structural Materials /PCB	ABS filament, acrylic, glass	\$76.79
Testing Materials	CO <sub>2</sub> and media	\$13.94

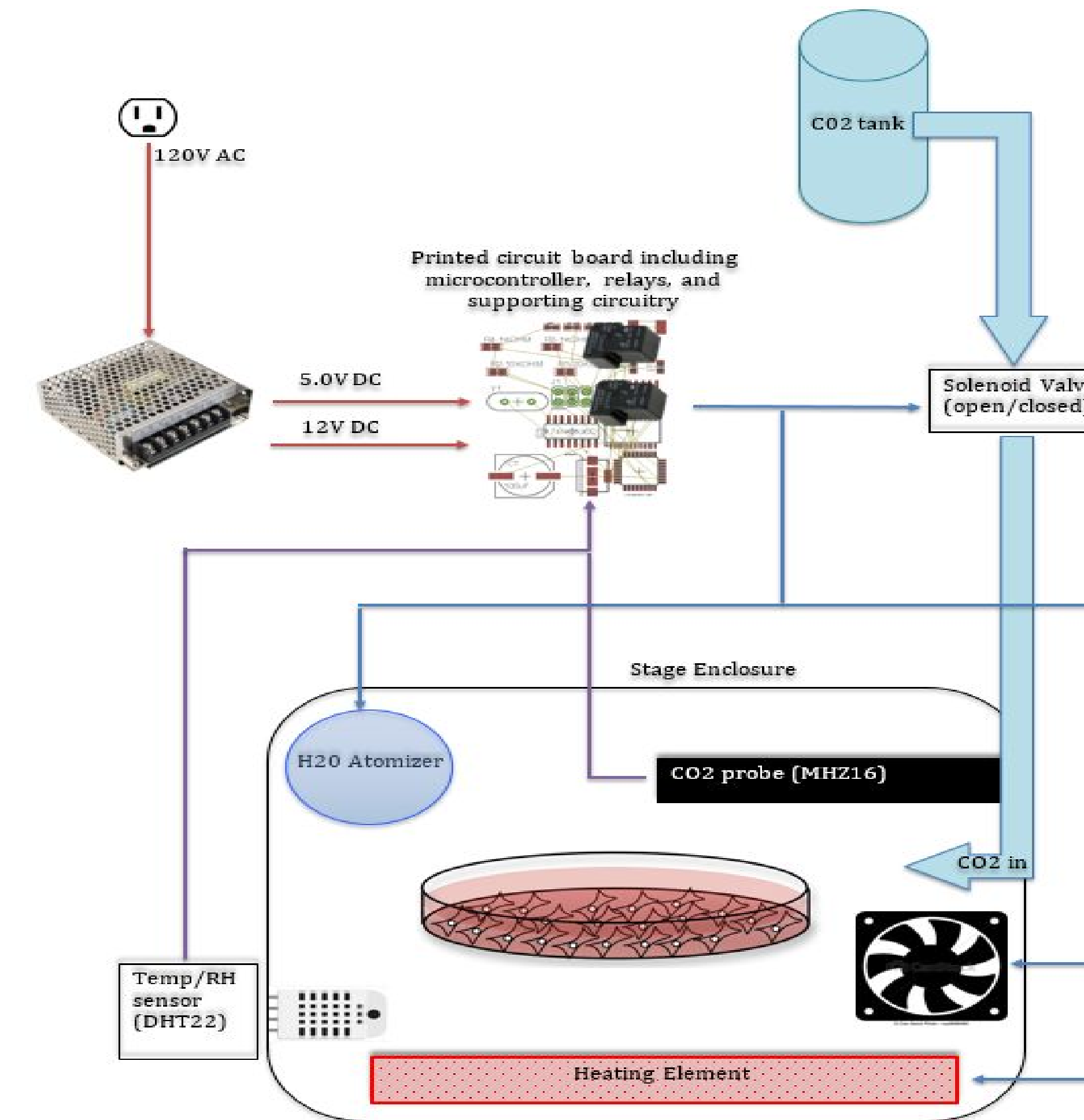


Figure 4. System diagram of final incubation system

## METHODS & TESTING

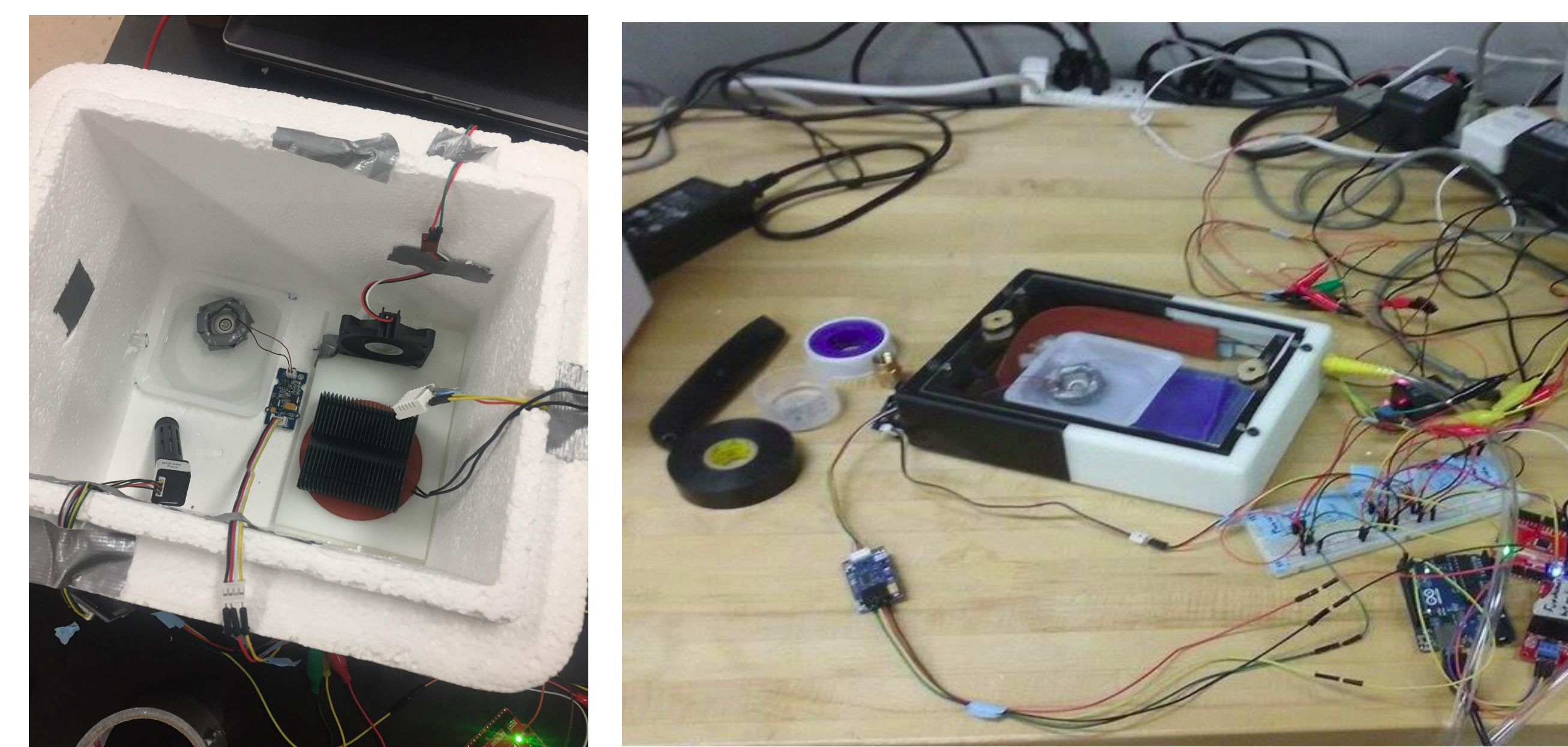


Figure 5. First iteration of control system validation (left) and second generation validation in fabricated enclosure(right)

- Cell Imaging Tests
  - Captured cell images in updated prototype
  - Focus achieved similar to control
- Cell Culture Tests
  - Scratch assay
    - test for migration
    - healthy cells migrate
  - Standard incubator
  - Final incubator prototype
  - Ambient conditions

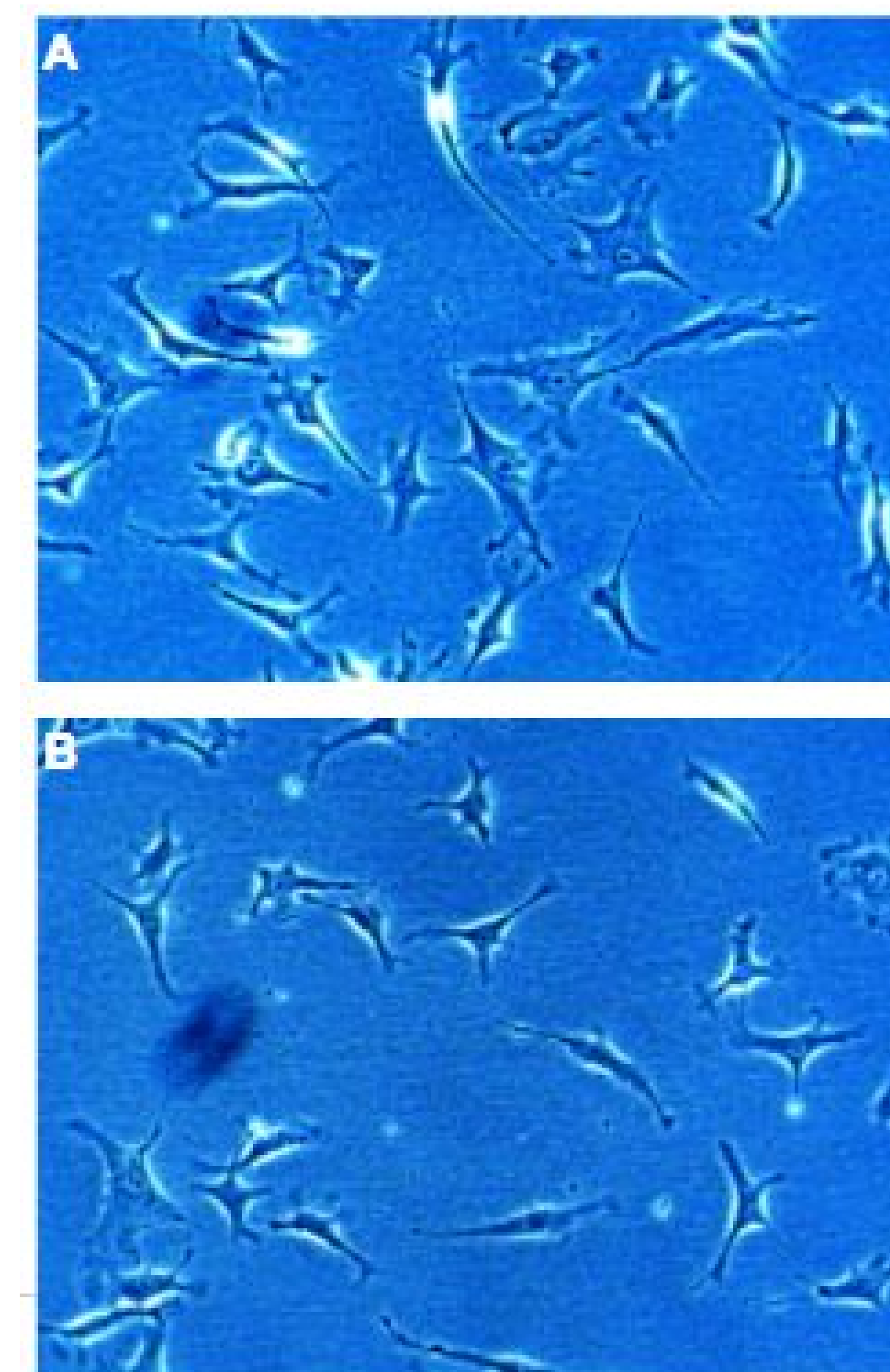


Figure 6. Images of cells in A) incubator, and B) on TCPS control at 20X magnification. Focus measure A = 31.8641% B = 31.0734%.

- Past semester testing:
  - Environmental feedback systems
    - Sensors and control circuitry for each parameter
    - Integration into one system
- Current semester testing:
  - Testing of integrated power supply and PCB
  - longer-term integrated testing

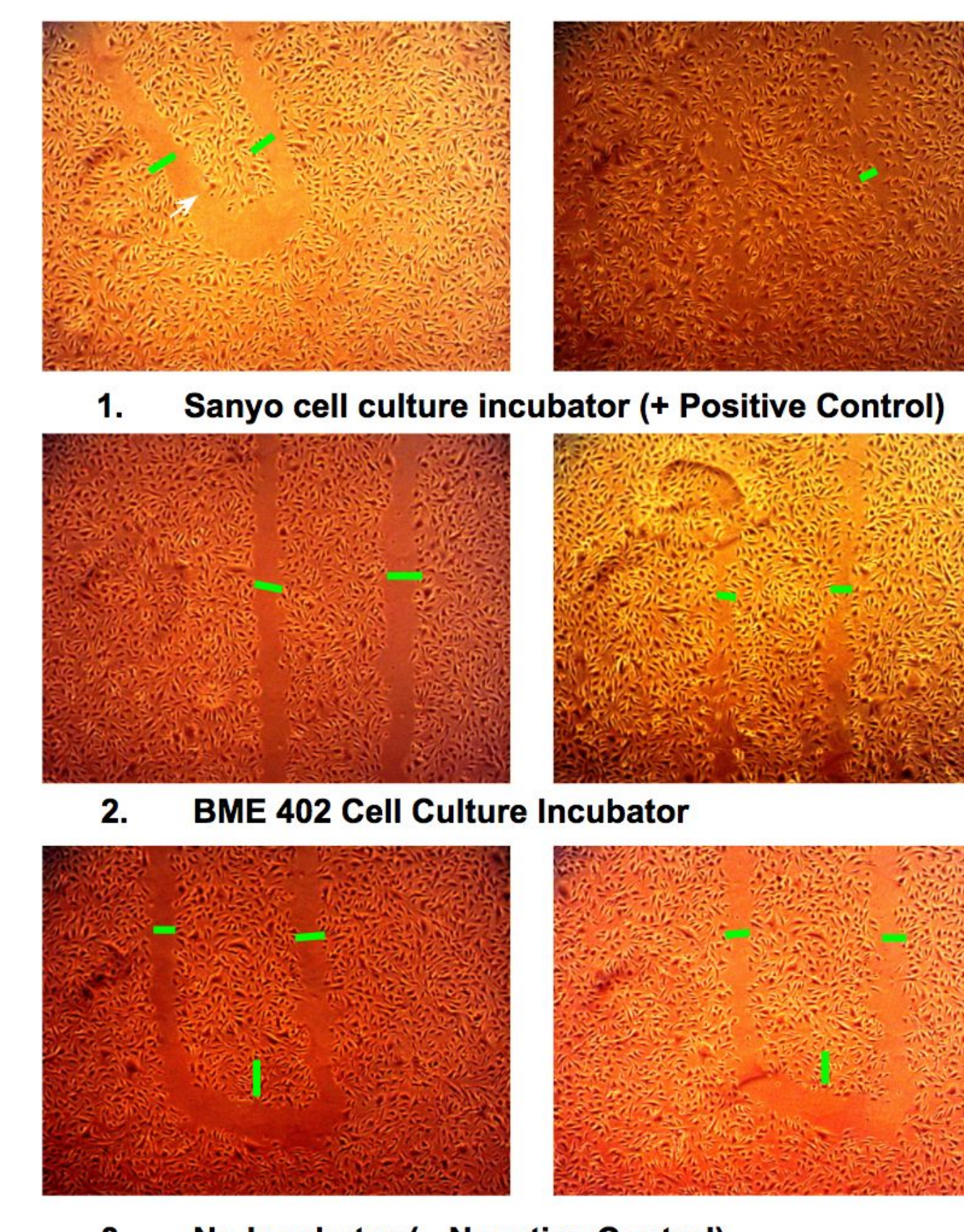


Figure 7. Images of cells migration in A) standard incubator, B) final prototype, and C) in ambient conditions. Lines added to show scratch "healing".

## RESULTS

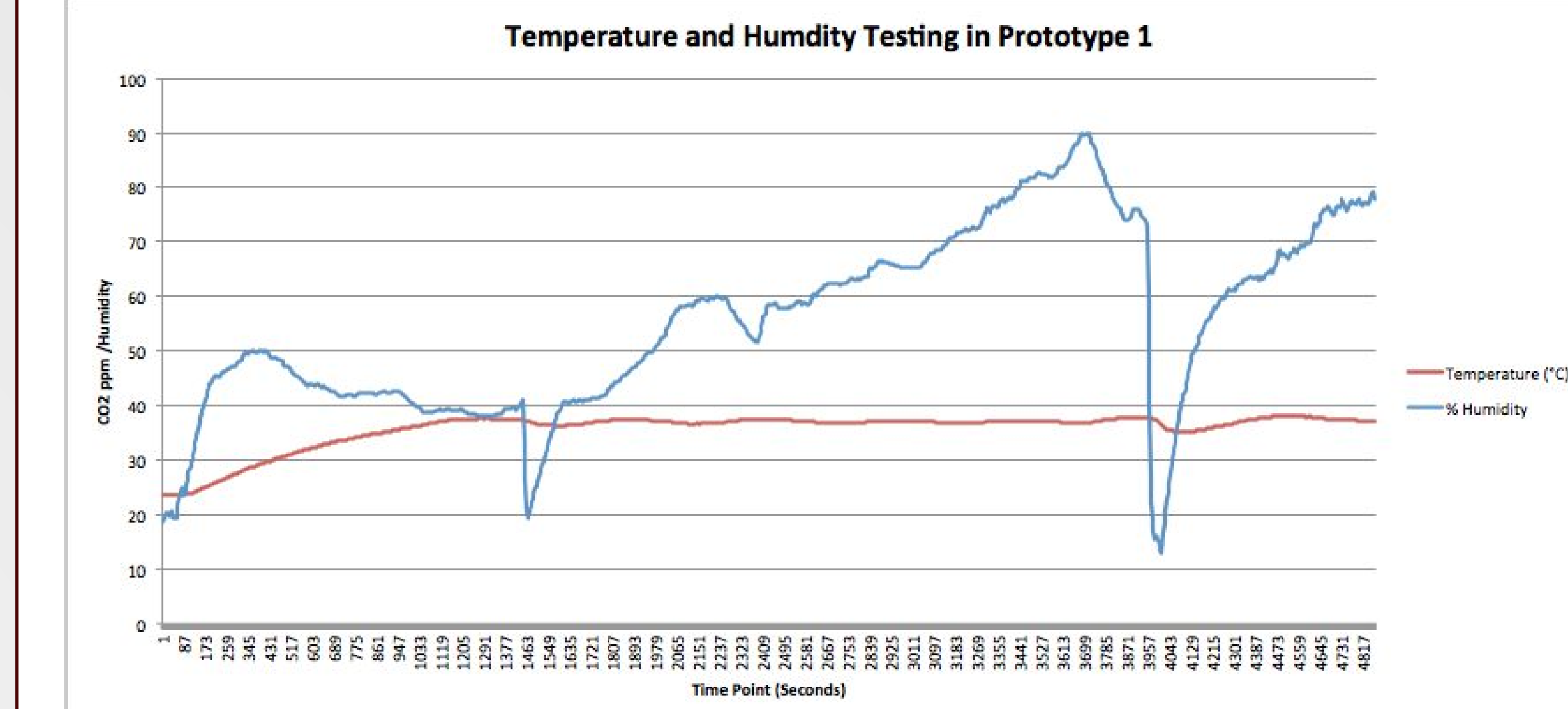


Figure 8. Humidity and Temperature data from first semester prototype.

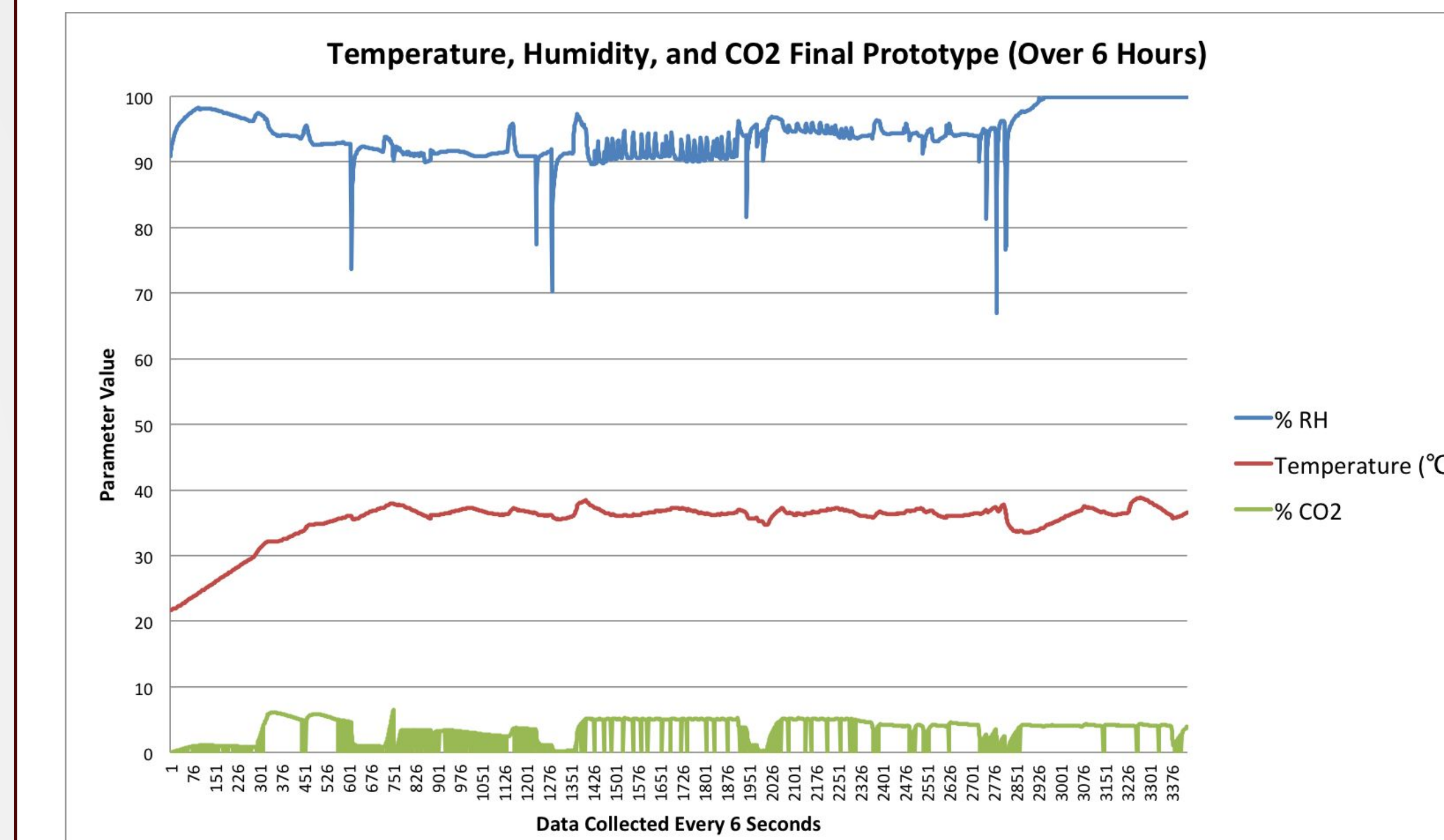


Figure 9. Humidity, temperature, and CO<sub>2</sub> data from final prototype.

## IMPACT & FUTURE WORK

- Extended testing duration with variety of cell types
- Further optimization of environmental control
- Selection of sensors/components for moving to larger volume production
- Design changes for manufacturing
  - Injection molded outer casing
  - Robust materials for sterilization

## ACKNOWLEDGEMENTS

- Dr. John Puccinelli
- Professor Mitch Tyler
- Dr. Amit Nimunkar
- Sam Lines



## REFERENCES

1. Baker, M (2010). "Cellular imaging: Taking a long, hard look". Nature. 466 (26): 1137-1140. doi:10.1038/4661137a.
2. "Miniature incubator for slides and petri dishes". Biosciencetools.com, 2016. [Online]. Available: [http://www.biosciencetools.com/catalog/incubator\\_Universal.htm](http://www.biosciencetools.com/catalog/incubator_Universal.htm). [Accessed: 17- Oct- 2016].
3. BioTek Imaging. "BioTek Imaging and Microscopy." BioTek Imaging and Microscopy, 2016. [Online]. Available: <http://www.cellimager.com/>. [Accessed: 19- Oct- 2016].
4. [http://www.pecon.biz/?page\\_id=3618](http://www.pecon.biz/?page_id=3618)
5. <http://www.pecon.biz/>