



DEPARTMENT OF
Biomedical Engineering
UNIVERSITY OF WISCONSIN-MADISON

Improving the precision of small human tissue biopsy processing

February 6, 2026

Team Members

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Overview of Presentation

- Client Description
- Problem Statement
- Broader Impact
- Design Constraints
- Fall 2025 Accomplishments
- Specific Goals & Timeline
- Packaging
- Documentation
- Budget



Client Description

- **Angela Gibson, MD, PhD, FACS**
- Specialties
 - General Surgery
 - Surgical Critical Care
 - Trauma Surgery
 - Burn & Wound Healing
- Performs research on epithelial regeneration of a burn injury and how to treat / expedite the healing process



Figure 1: Angela Gibson,
MD, PhD, FACS [1]



Figure 2: Bailey A. Donahue,
BS
Lab Technician/Manager [1]



Problem Statement

- **Objective:** fabricate a precise cutting device to shear fat off burn biopsy samples
- Gibson's lab creates burns in pig skin, to biopsy, and culture in media
 - Biopsies include, epidermis, dermis, and subcutaneous tissue
- Fat creates a hydrophobic layer that inhibits media absorption
 - Negatively affecting burn healing / tissue viability
- Current method is tedious, variable, and possibly damaging
 - Involves securing the sample with forceps and slicing with a scalpel

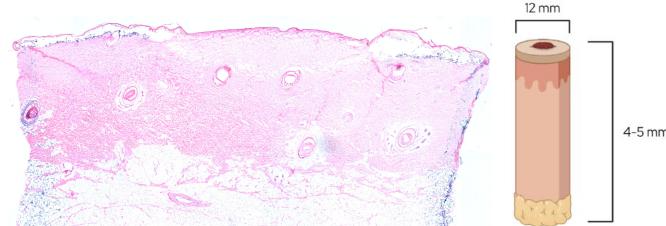


Figure 3: LDH stained pig skin sample *without* additional fat removal; lack of stain indicating poor viability [Bailey Donahue]

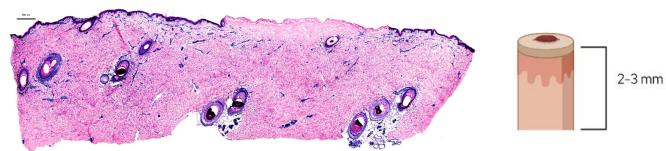


Figure 4: LDH stained pig skin sample *with* additional fat removal; blue LDH stain indicating viability [Bailey Donahue]

Motivation and Impact

- Burn injuries are the **leading cause of morbidity** worldwide [2]
- The Gibson Lab analyzes comparative pig skin models for **burn healing research** to better morbidity and mortality rates of burns [1]
 - Working to develop novel autologous burn wound regeneration technologies
- Removing additional fat from skin biopsy samples significantly **improves sample viability** and analysis capability
- Existing tissue slicing devices are not suitable for this process
 - High cost
 - Little adaptability

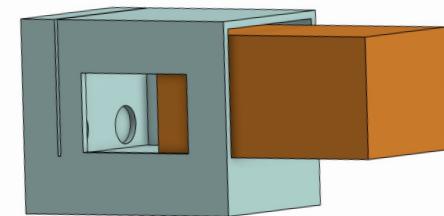
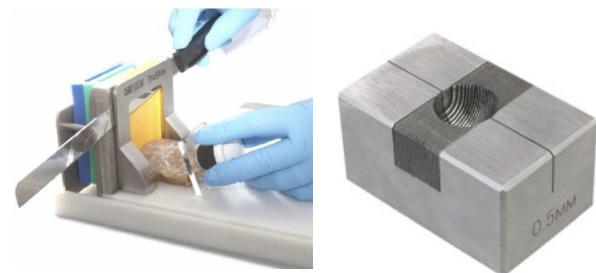


Figure 5: Client prototype OnShape assembly
[Bailey Donahue]



Figures 6 & 7: Tissue Slicing Systems from TedPella Inc. \$1878.75 and \$696-\$3630 [3][4]

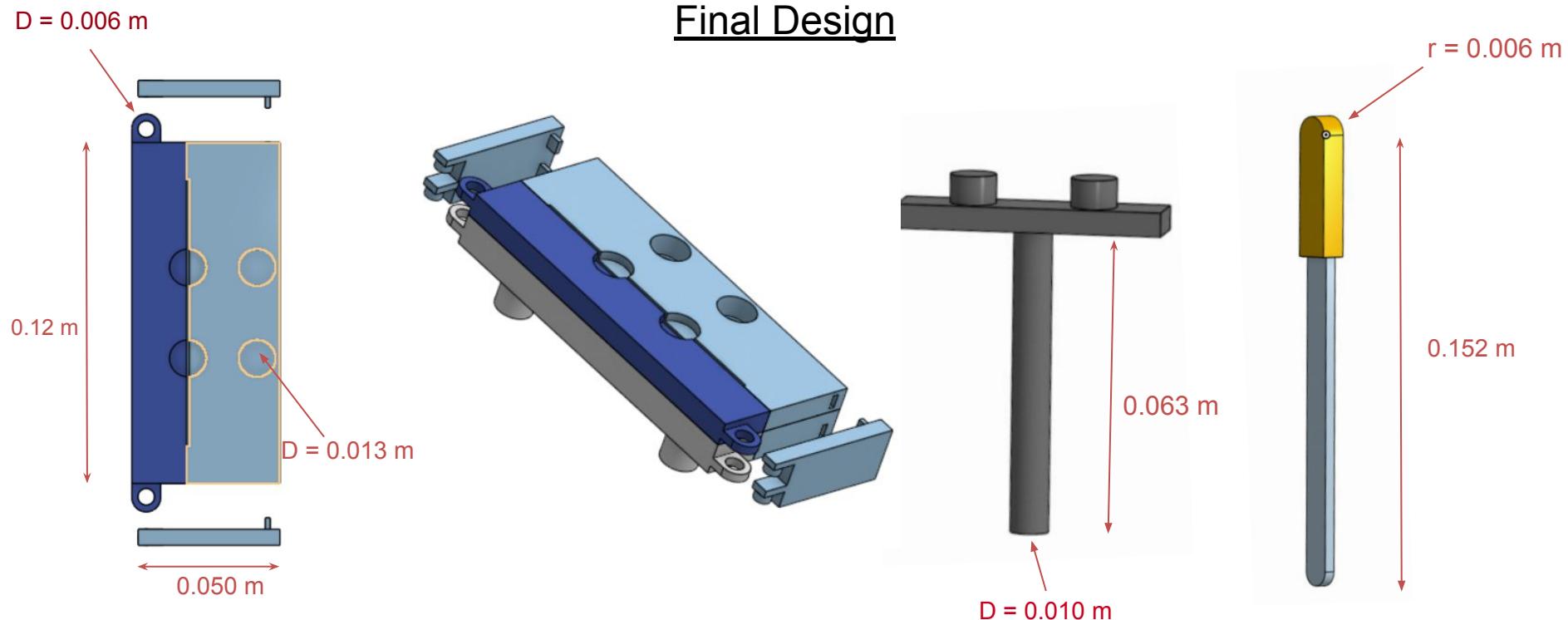


Design Constraints

- **Ease of Use:** Intuitive to use with a written protocol
- **Cut Accuracy & Precision:** Variability of the cut must be within $2.5 \pm .2$ mm
 - Straight cut
- **Maintenance:** Cleaned and sterilized with minimal material breakdown
 - Alcohol and bleach solutions
 - Autoclavable
- **Security of Biopsy:** Sample should be contained within the device while in use
 - Minimal movement
- **Ease of Fabrication:** 3D printable for replacement
- **Safety:** Minimize risk of injury to the user



Fall 2025 Accomplishments



Figures 8-11: OnShape drawings of the biopsy press, applicator, and blade handle

Fall 2025 Accomplishments

Final Design



Figure 12: Assembly of the rubber mat, biopsy press, blade handle, and tissue sample

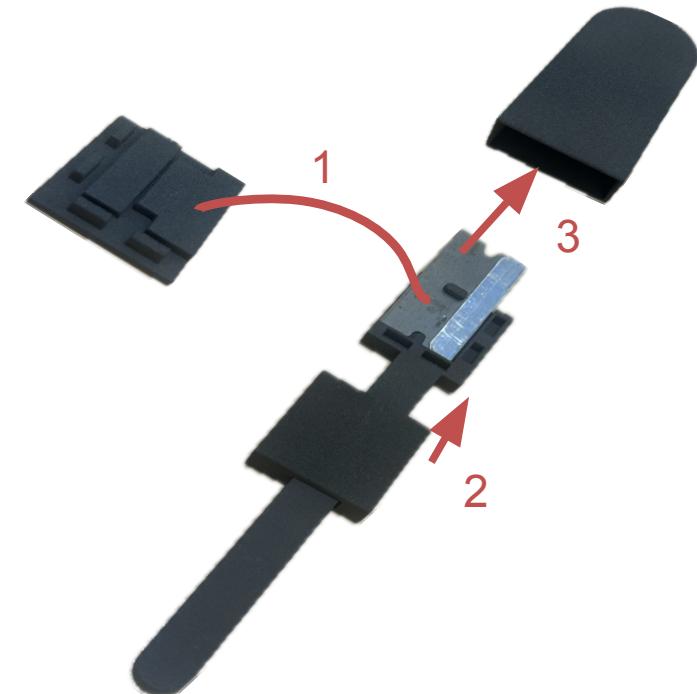


Figure 13: Assembly of the blade handle



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Testing & Results

- **Sanitization**
 - Residual fat in connectors & edges
 - Soap and water more effective than ethanol
- **Usability**
 - Yielded > 80% positive experiences in intuitiveness, device security, blade handle safety, thickness consistency, & overall effectiveness
- **FEA Simulation**
 - von Mises test applied substantial forces to the press and blade handle (>22N, >9N, respectively); no risk of deformation

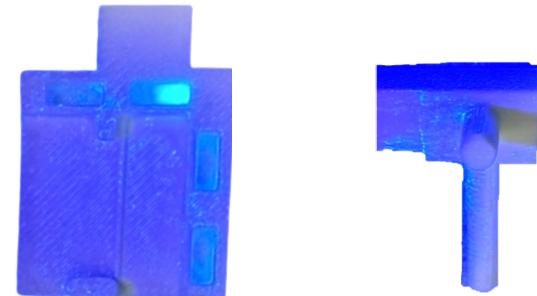


Figure 14: Sanitization test with UV light

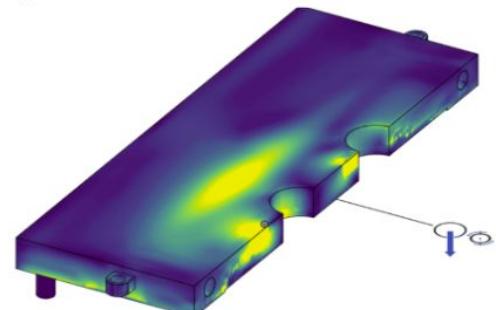


Figure 15: FEA simulation results [MPa]



Fall 2025 Accomplishments

Lessons Learned

- Nylon 12 (~\$20) vs PLA (~\$2) is a better option for longevity and demonstrates improved tissue preservation
- Overall design performs well
 - The blade fits in the cutting track and slice the sample evenly
 - FEA analysis validates the structural integrity of the biopsy press and handle
- Area for improvement: the pressure applicator
 - Hinders slicing when force is applied, leading to uneven cuts in the tissue



Specific Goals & Timeline

February



March



April

- Obtain more tissue (pig-skin) sample data from the clients (Feb. 20 & 27)
- Gather and analyze histology measurements received from clients
- Begin drafting the journal article
- Improvise the latest design with feedbacks and further consultations with clients
- Conduct more testing with new design to test compatibility with #11 scalpel blade
- Process the IP in design and patent process
- Revise the draft of journal article
- Final Poster Presentation
- Finalize & submit the journal article
- Deliver the final design to client



Packaging

- Autoclave sterilization
 - Pieces must be disassembled [5]
- Tyvek sterilization pouches [6]
- Tray [7]
 - Plastic
 - Aluminum
- Sterilizable wraps [8]
 - SMS



Figure 16: Sterilization wrapping of large medical devices [9]



Figure 17: Plastic Sterilizable Tray Case [10]



Figure 18: Tyvek sterilization pouches [11]



Documentation

User Manual



- Sample insertion
- Sample containment
- Blade insertion into cutline
- Cutting best practices
- Safety Warning - For the blade!

Sanitization instructions

Nylon 12 [12]:

- Alcohol resistance
- Ethylene oxide and formaldehyde solution
- Autoclave

Biomed Clear Resin [13]:

- Autoclave
- E-beam, gamma, and ethylene oxide



Material Expenditures

- **Last Semester: \$129.15**
- Largest sources of previous spending
 - Nylon 3D printed prototypes
 - Rubber base
- **Expected spending for this semester: ~ \$200**
- Largest anticipated expenditures
 - Further 3D printing with Nylon
 - Biomed clear resin
- Client has expressed preference for longevity over affordability



References

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Questions?



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